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(54) Title: HYBRID GROWTH FACTORS

(57) Abstract

The present invention provides recombinant hematopoietic molecules comprising at least a portion of a first hematopoietic molecule having early myeloid differentiation activity and at least a portion of a second hematopoietic molecule having late myeloid differentiation activity. Nucleic acid molecules encoding such recombinant molecules, as well as pharmaceutical compositions comprising such recombinant factors are also disclosed.

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⁺ Any designation of "SU" has effect in the Russian Federation. It is not yet known whether any such designation has effect in other States of the former Soviet Union.

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HYBRID GROWTH FACTORS

BACKGROUND OF THE INVENTION

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Within this application several publications are referenced by Arabic numerals within parentheses. Full citations for these references may be found at the end of the specification immediately preceding the Sequence Listing. The disclosures of these publications in their entirety are hereby incorporated by reference into this application in order to more fully describe the state of the art to which this invention pertains.

A variety of factors can influence the activity of a cell. Frequently a factor exerts its influence by interacting with a receptor on the surface of a cell. After binding to the receptor, the signal which determines the cellular response to the factor can be mediated through a number of different events, including internalization of the factor or alterations of the receptor caused by ligand binding. During the course of hematopoietic differentiation, a number of different factors are involved in the maturation of a pluripotent stem cell into a fully differentiated cell. The activities of these factors during the course of hematopoietic differentiation have resulted in these factors being characterized as early factors or late factors. For example, factors such as interleukin-3 (IL-3) and granulocyte-macrophage colony stimulating factor (GM-CSF) are considered early factors, while erythropoietin (Epo), macrophage colony stimulating factor (M-CSF), and granulocyte colony stimulating factor (G-CSF) are considered late factors.

Based upon studies performed with purified factors and <u>in vitro</u> colony forming unit assays, it appears that both IL-3 and GM-CSF act on pluripotent cells before they become committed to a particular hematopoietic pathway. After the events stimulated by these factors are underway, such lineage restricted cells become receptive to further differentiation mediated by such late factors as Epo, (which leads to the maturation of erythrocytes), G-CSF (which leads cells

into the granulocytic pathway), and M-CSF (which leads to the maturation of macrophages). Experiments described in recent publications (1,2,3) have demonstrated in vitro that early or late factors alone are poor stimuli of colony formation. However, when an early factor such as IL-3 or GM-CSF is combined with a late factor, levels of colony formation equivalent to that seen with conditioned media having full activity is observed. Thus, differentiation appears to be dependent upon the dual activities of early and late factors.

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Despite a clear requirement for both IL-3 or GM-CSF and Epo for the formation of erythroid colony forming units, published results indicate that IL-3 can down-modulate high affinity Epo receptors (4). Because the amount of IL-3 required to demonstrate down-modulation of the Epo receptor was higher than that reported by others who demonstrated functional full IL-3 activity in the presence of Epo, it is unclear whether this phenomenon is relevant in vivo.

Previous experiments in animals (22-26) suggest that under conditions of hematopoietic regeneration, optimal expansion of late progenitors could only occur in the presence of an adequate early progenitor pool. This then makes manipulations that result in the expansion of early hematopoietic progenitor pools extremely desirable. IL-3 has been shown to exert a differentiative and proliferative effect on early progenitor cells and at IL-3 concentrations which had little or no effect alone, Epo acted synergistically to induce proliferation and differentiation of erythroid progenitors. (27) By targeting a molecule with both early (IL-3) and late (Epo and G-CSF) activities to early progenitor cells, optimal expansion of a desired lineage should be possible.

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SUMMARY OF THE INVENTION

The present invention concerns hybrid molecules comprising early and late differentiation factors produced by genetic manipulation. By covalently linking such factors the local concentration of the late factor is very high at the surface of a cell to which the early factor is bound. Additionally, if down-modulation is relevant in vivo, binding of late factors to any remaining low-affinity receptors, e.g. Epo receptors, could be enhanced, thus reducing the amount of late factor required to stimulate the cell. Furthermore, by linking an early factor with a late factor, such early factor may act more specifically to stimulate only the desired lineage, thus reducing any undesirable effects mediated by the early factor. Finally, it is considerably easier to produce and administer to a patient a single factor with two activities rather it would to produce and administer two separate factors.

BRIEF DESCRIPTION OF THE FIGURES

Figure 1 shows a Western blot analysis of IL-3/Epo hybrid growth 20 factors in CHO CM. CHO CM was collected from clones 23-10 (IL-3:Epo Flex), 5-4 (IL-3:Epo Short) and 17-3-1 (Epo:IL-3 Short). growth factor concentrations were determined by ELISA assay. containing 74 ng of IL-3:Epo Flex (having a 23 aa flexible linker (lane 2), 73.5 ng of IL-3:Epo Short (having a short 2 aa linker) (lane 25 3), 80 ng of Epo:IL-3 Short (having a 3 aa linker) (lane 4) were subjected to SDS-PAGE (10-20% gel) electrophoresis and were assayed for Epo by Western blotting with a mouse anti-Epo polyclonal antisera Medium conditioned by CHO cells as described in Example 7. transfected with the vector pEe6 (lane 5) and rHu Epo 10 ng (lane 6), 30 20 ng (lane 7), 30 ng (lane 8), 70 ng (lane 9), and 100 ng (lane 10) were included. Molecular size markers in kilodaltons (lane 1).

Figure 2 shows AML193 cells proliferate in response to the IL-3 moiety of the hybrid growth factors. AML193 cells were grown to stationary phase and suspended in RPMI-1640 plus 10% FCS and growth

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factor deprived for 16 hours. The indicated concentrations of growth factors were added for 42 hours followed by a 6 hour pulse of (^3H) thymidine as described in Example 7. NO GF (no growth factor); CHO CM (medium conditioned by CHO cells transfected with the vector pEe6); Epo (rHu Epo); IL-3 (rHu IL-3); IL-3:Epo Flex (CHO CM containing IL-3:Epo fusion protein with a 23aa flexible linker); IL-3:Epo Short (CHO CM containing IL-3:Epo fusion protein with a 2aa linker); Epo:IL-3 Short (CHO CM containing Epo:IL-3 fusion protein with a 3aa linker).

Figure 3 shows dose response of IL-3 adapted AML193 cells to the IL-3 moiety of the hybrid growth factors. IL-3 adapted AML 193 cells were grown to stationary phase and suspended in RPMI-1640 plus 10% FCS minus growth factor for 16 hours. Increasing concentrations of IL-3 and fusion proteins were added and the assay was carried out as described in Figure 2 and in Example 7. IL-3:Epo Flex (CHO CM containing IL-3:Epo fusion protein with a 23aa flexible linker); IL-3:Epo Short (CHO CM containing IL-3:Epo fusion protein with a 2aa linker); Epo:IL-3 Short (CHO CM containing Epo:IL-3 fusion protein with a 3aa linker).

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Figure 4 shows FDC-P1/ER cells proliferate in response to the Epo moiety of the hybrid growth factors. FDC-P1/ER cells were grown to stationary phase and suspended in RPMI-1640 plus 10% FCS without growth factor for 16 hours. The indicated concentrations of growth factors were added for 42 hours followed by a 6 hour pulse of (³H) thymidine as described in Example 7. Columns are labeled as described in Figure 2. WEHI3 CM (medium conditioned by murine WEHI3 cells which produce and secrete IL-3).

Figure 5 shows dose response of FDC-P1/ER cells to the Epo moiety of the hybrid growth factors. FDC-P1/ER cells were grown to stationary phase and suspended in RPMI-1640 plus 10% FCS and deprived of growth factor for 16 hours. Increasing concentrations of Epo and fusion proteins were added and the assay was carried out as described in Example 7. Hybrid growth factors are as designated in Figure 3.

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Figure 6 shows IL-3 plus Epo responsiveness of IL-3 adapted TF-1 cells. TF-1 cells adapted for growth in IL-3 were grown to stationary phase and suspended in RPMI-1640 plus 10% FCS minus growth factor for 16 hours. Increasing concentrations of growth factors, 0.75 fmol/ml hybrid growth factors and 0.75 fmol/ml Epo plus 1.5 fmol/ml IL-3 (1), 1.5 fmol/ml hybrid growth factors and 1.5 fmol/ml of Epo plus 3.0 fmol/ml IL-3 (2), 3.0 fmol/ml hybrid growth factors and 3.0 fmol/ml Epo plus 6.0 fmol/ml IL-3 (3), were added and the assay was carried out as described in Example 7. Hybrid growth factors are as designated in Figure 3.

Figure 7 shows dose responsiveness of IL-3 adapted TF-1 cells to the hybrid growth factors. TF-1 cells adapted for growth in IL-3 were grown to log phase and suspended in RPMI-1640 plus 10% FCS minus growth factor for 16 hours. Increasing concentrations of hybrid growth factors were added and the cells were incubated for 8 hours. (3 H) Thymidine (1 μ Ci/well) was added and the incubation was continued for 16 hours. (A) Dose response to hybrid growth factor, concentrations of 0 to 30 fmol/ml. (B) Represents the same data as in A for concentrations of 0 to 1.875 fmol/ml to emphasize the differences between hybrid factors. Hybrid growth factors are as designated in Figure 3.

Figure 8 shows dose responsiveness of GM-CSF adapted TF-1 cells to the hybrid growth factors. TF-1 cells maintained in GM-CSF were grown to log phase and suspended in RPMI-1640 plus 10% FCS minus growth factor for 16 hours. Increasing concentrations of hybrid growth factors were added and the assay was carried out as described above for Figure 5. (A) Dose response to hybrid growth factor concentrations, of 0 to 30 fmol/ml. (B) Represents the same data as in A for concentrations of 0 to 1.875 fmol/ml to emphasize the differences between hybrid factors. Hybrid growth factors are as designated in Figure 3.

Figure 9 shows TF-1 cells proliferate in response to the IL-3 moiety of the IL-3/G-CSF hybrid growth factor. TF-1 cells were grown to stationary phase and suspended in RPMI-1640 plus 10% FCS deprived of

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growth factor for 16 hours. The indicated concentrations of growth factors were added and the assay was carried out as described in Example 7. Factors are as designated in Figure 2 except, G-CSF (rHu G-CSF); IL-3/G-CSF (CHO CM containing IL-3/G-CSF fusion protein with a 10aa linker).

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Figure 10 shows NFS-60 cells proliferate in response to the G-CSF moiety of the IL-3/G-CSF hybrid growth factor. NFS-60 cells were grown to stationary phase and suspended in RPMI-1640 plus 10% FCS minus growth factor for 16 hours. The indicated concentrations of growth factors were added and the assay was carried out as described in Example 7. Growth factors are as designated in Figures 2 and 9.

Figure 11 shows dose responsiveness of AML193 cells to the iL-3:G-CSF hybrid growth factor. AML193 cells were grown to stationary phase and suspended in RPMI-1640 plus 10% FCS deprived of growth factor for 16 hours. The indicated concentrations of growth factors were added and the assay was carried out as described in Example 7. Growth factors are as designated in Figures 2 and 9.

DETAILED DESCRIPTION OF THE INVENTION

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The present invention provides a recombinant hematopoietic molecule comprising at least a portion of a first hematopoietic molecule having early myeloid differentiation activity and at least a portion of a second hematopoietic molecule having late myeloid differentiation activity. This recombinant molecule has early myeloid differentiation activity associated with the first hematopoietic molecule and late myeloid differentiation activity associated with the second hematopoietic molecule. Within this application, "hematopoietic molecule" means a molecule which promotes and/or regulates hematopoiesis. Hematopoietic molecules exert such promotional or regulatory activities at different stages during hematopoiesis, such stages being referred to herein as early myeloid differentiation and late myeloid differentiation. Also within this application, "early myeloid differentiation activity" means the ability to promote the

differentiation, self-renewal, or proliferation of pluripotent myeloid cells, i.e., stem cells or colony forming unit, granulocyte-erythrocyte-monocyte-megacaryocyte, cells. Moreover, within this application, "late myeloid differentiation activity" means the ability to promote the maturation or differentiation of a lineage restricted myeloid cell, i.e., a myeloid precursor cell committed to a specific cell lineage such as erythrocytes, megakaryocytes, monocytes, neutrophils, eosinophils, and basophils.

In one embodiment of the invention, the first hematopoietic molecule is selected from the group consisting of IL-3 and GM-CSF. In another embodiment of the invention, the second hemopoietic molecule is selected from the group consisting of Epo, G-CSF, IL-5 and M-CSF. In a preferred embodiment of the invention, the portion of the first hematopoietic molecule is linked to the portion of the second hematopoietic molecule by an amino acid linker sequence comprising at least two amino acid residues.

Within the context of the present invention, it is understood that variations in proteins and nucleic acids exist among individuals, e.g. amino acid or nucleotide substitutions, deletions, insertions, and degree or location of glycosylation, and that functional derivatives resulting therefrom are included within the scope of the present invention.

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In a preferred embodiment of the invention, the recombinant molecule comprises the entire amino acid sequence of human IL-3 (SEQ ID NO: 1). Moreover, the recombinant hematopoietic molecule may preferably comprise a 79 amino acid sequence derived from human IL-3 (SEQ ID NO: 2), i.e. residues 1-79 of SEQ ID NO: 1.

Further still, in yet another preferred embodiment of the invention, the recombinant molecule comprises the entire amino acid sequence of human erythropoietin (SEQ ID NO: 3). In still a further embodiment of the invention, the hemopoietic molecule comprises a 155 amino acid

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sequence derived from human erythropoietin (SEQ ID NO: 4), i.e., residues 7-161 of SEQ ID NO: 3.

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In another preferred embodiment of the invention, the recombinant hematopoietic molecule comprises the entire amino acid sequence of human G-CSF (SEQ ID NO: 5).

In one embodiment of the invention, the first hematopoietic molecule is IL-3 and the second hematopoietic molecule is erythropoietin. The first hematopoietic molecule, i.e. IL-3, may comprise the amino portion and the second hematopoietic molecule, i.e. Epo, may comprise the carboxyl portion of the recombinant molecule. Preferably, the recombinant hematopoietic molecule comprises the amino acid sequence from amino acid 1 to amino acid 302 of SEQ ID NO: 6. Also preferably. the recombinant hematopoietic molecule comprises the amino acid sequence from amino acid 1 to amino acid 321 of SEO ID NO: 7. However, in another embodiment of the invention, the first hematopoietic molecule, i.e. IL-3, may comprise the carboxyl portion and the second hemopoietic molecule, i.e. Epo, may comprise the amino portion of the recombinant molecule. In a preferred embodiment of the invention, the recombinant molecule comprises the amino acid sequence from amino acid 1 to amino acid 303 of SEO ID NO: 8. In yet another preferred embodiment, the recombinant molecule comprises the amino acid sequence from amino acid 1 to amino acid 322 of SEQ ID NO: 9.

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In still a further embodiment of the invention, the first hematopoietic molecule is IL-3 and the second hematopoietic molecule is G-CSF. In one such embodiment, the first hematopoietic molecule comprises the amino portion and the second hematopoietic molecule comprises the carboxyl portion of the recombinant molecule. In yet a more specific embodiment, the recombinant molecule comprises the amino acid sequence from amino acid 1 to amino acid 317 of SEQ ID NO: 10.

The subject invention also provides nucleic acid molecules which encode the recombinant hematopoietic molecules of the subject invention. Examples of such nucleic acid molecules are SEQ ID NO: 11,

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SEQ ID NO: 12, SEQ ID NO: 13, SEQ ID NO: 14 and SEQ ID NO: 15. Moreover. vectors which comprise the nucleic acid molecules of the subject invention are also disclosed. In one embodiment of the invention, the vector comprises a plasmid. Moreover, host vector systems for the production of a recombinant hematopoietic molecule of the present invention are provided which comprise a vector of the present invention in a suitable host, preferably a mammalian cell such as a CHO or COS cell. This host vector system may be grown under suitable conditions which permit the expression of the recombinant hematopoietic molecule, which may be recovered by purification techniques known in the art, e.g. ion exchange chromatography, affinity chromatography, and size exclusion chromatography.

The present invention further provides pharmaceutical compositions useful for treating patients suffering from anemias of various origins, e.g. renal failure, and AIDS. Moreover, these pharmaceutical compositions are useful for administering to patients for preoperative autologous blood donations, patients receiving or donating bone marrow for transplantation purposes, and patients undergoing cancer chemotherapy. These pharmaceutical compositions comprise effective hematopoiesis-promoting amounts of a recombinant molecule of the present invention and a pharmaceutically acceptable carrier. Pharmaceutically acceptable carriers are known in the art and are disclosed in The Pharmacopeia of the United States and the National Formulary. Depending on the specific application contemplated, the pharmaceutical composition may be formulated as a solution, suspension, parenteral preparation, or spray. Parenteral preparations may include a vehicle such as specially distilled, pyrogen-free water, phosphate buffer, or normal saline. Oral and/or transmucosal dosage forms may comprise phospholipids, often in the form of liposomes.

Also provided is a method for treating a patient to promote hematopoiesis which comprises administering to the patient an effective hematopoiesis-promoting amount of a pharmaceutical composition of the present invention.

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The recombinant hematopoietic molecules, nucleic acid molecules, pharmaceutical compositions and methods of the present invention will be better understood by reference to the following experiments and examples, which are provided for purposes of illustration and are not to be construed as in any way limiting the scope of the invention, which is defined by the claims appended hereto.

Examples

Construction of the hybrid protein genes: Genes encoding IL-3 (SEQ ID NO: 16), Epo (SEQ ID NO: 17) and G-CSF (SEQ ID NO: 18) were purchased from British Biotech. Ltd. These genes were utilized to construct three different hybrid hematopoietic proteins, i.e., IL-3:Epo, Epo:IL-3 and IL-3:G-CSF. In these hybrids the first named gene forms the amino portion and the second named gene the carboxyl portion of the hybrid protein.

Example 1

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A nucleic acid molecule encoding an IL-3:Epo hybrid growth factor was constructed as follows: CSF, the native leader sequence of IL-3 was synthesized as 4 oligonucleotides (SEQ ID NOS: 19-22; see Table I) which represents both strands of the leader sequence. In addition, the 5' end of the leader (SEQ ID NO: 19) encoded a convenient restriction enzyme overhang (EcoR1), although the EcoR1 site was not regenerated, in front of the ATG start codon. The 3' end of the leader (SEQ ID NO: 21) included the first several amino acid codons of IL-3 and an SpeI overhang so that the annealed leader sequence could be easily ligated to IL-3, which was altered by British Biotech to include an SpeI site. The leader sequence was annealed and ligated to pKS (Stratagene Cloning Systems, Inc., San Diego, CA) cleaved with EcoRl and SpeI. The resulting plasmid was designated pKSO. The IL-3 containing pUC18 plasmid obtained from British Biotech was cleaved with SpeI and NheI, then ligated to a linker oligonucleotide (complimentary oligonucleotide SEQ ID NOS: 23 and 24; see Table I) which contained the following three restriction sites: NheI, XbaI and

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NcoI. Cleavage was then performed with SpeI and XbaI. The resulting 379 base pair fragment was then ligated to PKSO cleaved with

SpeI and XbaI. The resulting plasmid (pKSOIL-a) contained the IL-3 leader, the IL-3 gene and a small linker fragment.

The Epo gene was inserted into pEe6 (Celltech, Ltd., Slough, U.K.), a mammalian expression vector which contains the human Cytomegalovirus promoter, a polylinker region and a poly-A addition site in addition to ampicillin resistance and a bacterial origin of replication, by cleaving the Epo containing plasmid obtained from British Biotech with HindIII and BamHl. Epo was then cleaved with NcoI. The same linker comprising oligonucleotide SEQ ID NOS: 23 and 24 as described earlier was ligated to Epo and then cleaved with XbaI to yield the entire Epo gene. This was then ligated to XbaI and BclI cleaved pEe6 to yield pEe6 containing the Epo gene (pEepo). PKSOIL-a was cleaved with EcoRY and an XbaI linker was ligated to the blunt ends followed by cleavage with XbaI, which released the IL-3 gene with the leader sequence. This was then ligated to XbaI cleaved Peepo to yield a plasmid containing an entire hybrid protein gene (pEepie-a) (see SEQ ID NO: 11 for the structure of the inserted hybrid gene, designated herein IL-3:Epo Short). The glutamine synthetase (gs) gene was then inserted into the BamH1 site of pEepie-a to yield pEepogs-a or pEpogs-b, depending upon the orientation of the gs gene. Glutamine synthetase confers resistance to methionine sulphoximine (MSX) in order to select cells which have taken up the plasmid after transfection. After the plasmid was constructed a large batch was grown, purified by CsCl ultracentrifugation, and used for transfection. At each step in this process all ligation joints between fragments were analyzed by DNA sequence analysis in order to assure that there were no changes that would cause frameshifts and prevent the hybrid gene from being expressed.

To construct a nucleic acid molecule encoding an IL-3:Epo hybrid growth factor with a longer linker sequence separating IL-3 and Epo, pEepie-a was cleaved with NheI and annealed oligonucleotide SEQ ID

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NOS: 25 and 26 (see Table I) were ligated into the cleaved plasmid. This linker encodes the flexible amino acid sequence Gly Ser Gly Ser Gly Ser Gly Ser (SEQ ID NO: 27). Clones with the insert in the proper orientation were selected by probing colonies with the junction oligonucleotide SEQ ID NO: 28 (see SEQ ID NO: 14 for the structure of the inserted hybrid gene, designated herein IL-3:Epo Flex). The glutamine synthetase gene was then added to the construct as described above.

10 Example 2

A nucleic acid molecule encoding an IL-3:G-CSF hybrid growth factor was constructed as follows: pUC18 containing G-CSF (British Biotech) was cleaved with HindIII. A linker composed of an overhanging XbaI site, a NotI site and an overhanging HindIII site (oligonucleotide SEQ ID NOS: 29 and 30; see Table I) was ligated to the pUC18:G-CSF. This was then cleaved with XbaI and BamHl which released the entire G-CSF gene. The G-CSF fragment was then inserted into XbaI and BclI cleaved pEe6 (pEe6:G-CSF). IL-3 with its signal sequence was removed from the IL-3:Epo plasmid pEepogs-a as an XbaI fragment. This IL-3 fragment was then inserted into XbaI cleaved pEe6-G-CSF. After restriction analysis, a plasmid containing the IL-3 gene in the proper orientation was obtained (pEG11), this plasmid encoded a gene capable of expressing IL-3 and G-CSF as a hybrid protein (see SEQ ID NO: 13 for the structure of the inserted hybrid gene, designated herein IL-3:G-CSF). The gs gene was inserted into this plasmid as described in Example 1 above to yield plasmids pEG13 and pEG14, depending upon the orientation of the qs gene.

30 <u>Example 3</u>

A nucleic acid molecule encoding an Epo:IL-3 hybrid growth factor was constructed by first synthesizing the native Epo signal sequence as oligonucleotide SEQ ID NOS: 31-36 (see Table I). These were annealed to yield an overhanging 5' XhoI sequence and a 3' PstI sequence. These were then ligated and subcloned as an XhoI/PstI fragment

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(pEpol). In order to obtain the proper reading frame and signal sequence processing site, the plasmid containing the signal sequence was cleaved with PstI and the 3' overhang left by PstI was enzymatically removed with T4 polymerase. This was then cleaved with BamHl. The Epo gene was then amplified by PCR as a fragment with a 5' blunt end using oligonucleotide SEO ID NO: 37 as a primer and a 3' BamHl end using oligonucleotide SEQ ID NO: 38 as a primer. This fragment was then ligated into pEpol to yield a complete Epo gene with its leader sequence. PCR was used to amplify the Epo gene with its signal sequence as an (5') XbaI and (3') NotI fragment using oligonucleotide SEQ ID NOS: 39 and 40 as primers. This was then digested with XbaI and NotI. At the same time, a purified IL-3 fragment was amplified by PCR as a (5'! NotI and (3') BamHl fragment using oligonucleotide SEQ ID NOS: 41 and 42, followed by digestion with NotI and BamHl. These two fragments were ligated to pEe6 cleaved with Xbal and BclI to yield a full length hybrid gene encoding both Epo and IL-3 (pEG16) (see SEQ ID NO: 12 for the structure of the inserted hybrid gene, designated herein Epo:IL-3 Short). The gs gene was inserted as described in Example 1 above to yield pEG17 and pEG18, depending upon the orientation of the gs gene.

A flexible linker is inserted into Epo:IL-3 by cleaving pEG17 or pEG18 with NotI. Annealed oligonucleotide SEQ ID NOS: 43 and 44 are then ligated into the cleaved plasmid. Clones with the insert in the proper orientation are selected by probing colonies with a junction oligonucleotide as described above (see SEQ ID NO: 15 for the structure of the inserted hybrid gene.)

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TABLE I

<u>OLIGONUCLEOTIDES</u>

All oligonucleotides are listed in the 5' to 3' orientation:

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       AATTGCCGCC ACCATGAGCC GCCTGCCCGT CCTGCTCCT (SEQ ID NO: 19)
       GCTCCAACTC CTGGTCCGCC CCGGACTCCA AGCTCCCATG ACCCAGACAA (SEQ ID NO: 20)
       CTAGTTGTCT GGGTCATGLG AGCTTGGAGT CCGGGGCGG (SEQ ID NO: 21)
       ACCAGGAGTT GGAGCAGGAG CAGGACGGGC AGGCGGCTCAT GGTGGCGGC (SEQ ID NO: 22)
       CTAGCGATCT TTCTAGA (SEQ ID NO: 23)
       CATGTCTAGA AAGATCG (SEQ ID NO: 24)
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       CTAGAAGCGG CCGCA (SEQ ID NO: 29)
       TTCGCCGGCG TTCGA (SEQ ID NO: 30)
       TCGAGCCATG GGGGTGCACG AATGTCCT (SEQ ID NO: 31)
       GCCTGGCTGT GGCTTCTCCT GTCCCTGCTG TC (SEO ID NO: 32)
       GCTCCCTCTG GGCCTCCCAG TCCTGGGCTG CA (SEQ ID NO: 33)
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       GCCCAGGACT-GGGAGGCCCA GAGGGA (SEQ ID NO: 34)
       GCGACAGCAG GGACAGGAGA AGCCACAGCC AGGCAGGACA TT (SEQ ID NO: 35)
      CGTGCACCCC CATGGC (SEQ ID NO: 36)
       GCCCCACCAC GCCTCATCTG T (SEQ ID NO: 37)
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       GAATTCGGAT CCTTATCATC T (SEQ ID NO: 38)
       CTAGTCTCTA GAATGGGGGT CCACGAATGT (SEQ ID NO: 39)
      AGCCATGGCG GCCGCTCTGT CCCCTGTCCT (SEQ ID NO: 40)
       GACAGAGCGG CCGCCATGGC TCCCATGACC (SEQ ID NO: 41)
       GAATTCGGAT CCTTACTAAA AGATCGCTAG (SEQ ID NO: 42)
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       CTAGCGTCCG GAGGCGGTGG CTCGGGCGGT GGCGGCTCGG GTGGCGGCG GCTCTGCG
       (SEQ ID NO: 25)
       CTAGCGCAGA GCCGCCGCCA CCGCAGCCGC CACCGCCCGA GCCACCGCC TCCGGACG
       (SEQ ID NO: 26)
       TTGTCGCTAG CGTCCGGAGG C (SEQ ID NO: 28)
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       GGCCGCTTCC GGAGGCGGTG GCTCGGGCGG TGGCGGCTCG GGTGGCGGC GGCTCTGC
       (SEQ ID NO: 43)
       GGCCGCAGAG CCGCCGCCAC CCGAGCCGCC ACCGCCCGAG CCACCGCCT CCGGCAGC
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(SEQ ID NO: 44)

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Example 4

Transfection of the hybrid gene containing plasmids. A11 transfections were performed using the Lipofectinç transfection kit (Bethesda Research Labs, Gaithersburg, MD) using 15-30 mg. of purified plasmid DNA (pEepogs-a, pEepogs-b, pEG13, pEG14, pEG17, and pEG18). The following alterations were made to the protocol provided by the company: the growth medium in these experiments was GMEM-S and the CHO-K1 cells were incubated in the presence of 10% CO₂; after addition of the lipofectin: DNA complex, cells were incubated without selection for 24 hours. The cells were transferred to GMEM-S supplemented with 25 mM MSX after 24 hours. The MSX concentration was subsequently Cloning rings were used to increased to 50 mM after one week. subclone MSX resistant colonies and each of these colonies was placed into an individual well of a 24 well plate. Selected clones were incubated in the absence of MSX to insure that the hybrid protein gene was stably integrated. Strongly positive clones were grown in large cultures to provide larger amounts of hybrid proteins for further analysis.

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Example 5

Assays for hybrid protein production. Cell supernatants from transfected or control cells were assayed using several different assays. In order to demonstrate Epo production, an RIA kit for Epo was used (Incstar Corp., Stillwater, MN). The presence of IL-3 was determined using an ELISA assay in which the capture antibody was a polyclonal goat anti-IL-3 (R&D Systems, Minneapolis, MN) and the probe antibody was a murine anti-IL-3 monoclonal. Goat anti-mouse conjugated to horseradish peroxidase followed by suitable substrate was used to detect the presence of the monoclonal anti-IL-3. A very similar assay was used to demonstrate the presence of the hybrid proteins except that a murine anti-Epo monoclonal or anti-G-CSF monoclonal was used in place of anti-IL-3 monoclonal. Additionally, IL-3:Epo Short was analyzed by Western blot analysis. The blot was probed with antibody to Epo and then with $^{125}\mathrm{I}$ goat anti-mouse. A

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single broad band appeared on the autoradiogram with a molecular weight of slightly more than 50,000 daltons.

Example 6

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Celiular assays. Epo and/or IL-3 dependent and responsive cell lines were used to test the biological activities of the hybrid proteins. B6SUtA (5) is a multipotential hematopoietic progenitor cell line established from nonadherent cell populations removed from continuous 86.S mouse bone marrow culture. This cell line demonstrates absolute dependence upon a source of growth factor(s). In response to Epo a population of the cells synthesize hemoglobin. Studies of globin expression indicated that the globin programs of B6SUtA cells are similar to those of erythroid progenitors at the period of transition from the yolk sac to fetal liver erythropoiesis. TF-1 (6) it is a cell line of immature erythroid origin established from a patient with erythroleukemia. The cell line shows complete dependency on GM-CSF or Epo sustains short-term growth of TF-1 and will induce IL-3. hemoglobin synthesis in a very small population of cells (8%). Hemin and w-aminolevulinic acid induce hemoglobin synthesis in most of the cells.

Human IL-3 will not bind the murine IL-3 receptor, therefore experiments that were done with B6SUtA cells measured only the functionality of the Epo moiety of the hybrid. B6SUtA cells are carried in murine IL-3. In each experiment, they are washed thoroughly and set up with growth factors at 10⁵ cells/ml. Cell growth and hemoglobin content were monitored on days 3 and 6 of each experiment. Cells grown in the presence of concentrated (10X) CH0 conditioned medium (CM) containing IL-3:Epo Short at a final concentration equivalent to 4.8 units/ml of Epo grew as well as cells grown in an equivalent amount of recombinant human (rHu) Epo. The percentage of cells which synthesized hemoglobin in response to the CHO-IL-3:Epo Short CM was always four times that of cells exposed to rHu Epo. B6SUtA cells grown in the presence of rHu IL-3 and rHu Epo grew as well as cells grown in the presence of IL-3:Epo Short and

induced hemoglobin synthesis in the same percentage of cells as did rHu Epo. Cells exposed to recombinant murine IL-3 (rMu IL-3) and rHu Epo grew similarly to cells exposed to rMu IL-3 alone and neither effectively induced cells to synthesize hemoglobin. Concentrated control CHO CM did not support the growth of B6SUtA cells nor did it induce hemoglobin synthesis. CHO CM plus rHu Epo supported cell growth and hemoglobinization as well as CHO-IL-3:Epo Short CM.

CHO-IL-3:Epo Short CM as well as CHO-rHu IL-3 CM both supported growth of human TF-1 cells. Control CHO CM supported only limited growth of the TF-1 cells.

Discussion

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The above-mentioned results demonstrate that a hybrid protein comprising two growth factors can be expressed in mammalian cell culture systems. In vitro assays of IL-3:Epo Short indicate that this hybrid protein has the activities of both IL-3 and Epo. The therapeutic application of such hybrid factors has advantages over using two factors separately simply in terms of patient administration, and moreover since the production, purification and formulation of one factor is less labor intensive than for two separate factors.

25 Example 7

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Factor Dependent Cell Lines and Culture Media - The GM-CSF/IL-3/Epo dependent human TF-1 cell line and the G-CSF dependent murine NFS-60 cell line were grown and maintained as described (7,8,). The GM-CSF dependent human cell line AML 193 (9) was adapted for growth in IL-3 by continuous culture of the cells in RPMI-1640 plus 10% FCS supplemented with rHu IL-3 for 6 weeks. The TF-1 derived cell line, TF-136 was selected by continuous culture of the TF-1 line in RPMI-1640 plus 10% FCS supplemented with 5ng/ml of rHu IL-3 for 6 months, followed by single cell suspension cloning of the resultant IL-3 dependent cells. The Epo dependent murine cell line, FDC-P1/ER,

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was derived from the IL-3 dependent line, FDC-P1, by introduction of the murine Epo receptor into these cells. (10) FDC-P1/ER cells are maintained in RPMI-1640 plus 10% FCS supplemented with 1 unit/ml of rHu Epo. Recombinant human Epo was obtained from Ortho Biologicals, Inc (Raritan, NJ). Recombinant human IL-3, rHu G-CSF and rHu GM-CSF were purchased from R & D Systems (Minneapolis, MN).

Capture ELISA Assay - ELISA plate was coated with 5 μ g/200 μ l/well of goat anti-human IL-3 (R & D Systems) in PBS at 40°C overnight. Excess antibody was removed by washing with PBS. Blocking was carried out with 300 μ l/well of 1% non-fat milk in PBS for 1 hour at 37°C followed by washing with 0.05% Tween^m in PBS. Samples were then incubated with the IL-3 antibody for 1 hour at 37°C in 0.5% non-fat milk, 0.025% Tween^m. Following extensive washing, the second antibody, a mouse anti-Epo monoclonal (Genzyme, Cambridge, MA), was added to the plate which was incubated for 1 hour at 37°C. The plate was washed and incubated with conjugate antibody (Goat anti-mouse-horseradish peroxidase) for 30 minutes at 37°C. Color development was carried out with the addition of o-phenylenediamine/ H_2O_2 at room temperature (RT) for 30 minutes. The reaction was stopped with 1N H_2SO_4 and the samples were read at 495 nm.

Gene Amplification- CHO cell lines producing significant amounts of the hybrid growth factors were isolated and 10^6 cells were plated in a 75 mm T-flask in GMEM-S medium containing various concentrations of methionine sulphoximine (MSX), ranging between 100μ M and 500μ M. Colonies resistant to the highest MSX concentration (IL-3:Epo Flex 200μ M; IL-3:Epo Short 250μ M; Epo:IL-3 Short 250μ M; IL-3:G-CSF 250μ M) were isolated and expanded. Those clones producing the highest levels of hybrid growth factors as determined by Epo or G-CSF ELISA assay (Amgen) were used for subsequent studies. IL-3:Epo Flex (clone 23-10); IL-3:Epo Short (clone 5-4); Epo:IL-3 Short (17-3-1).

Cell Proliferation Assays - Factor dependent cells were grown to stationary phase, washed, and incubated for 16 hours in media plus 10% FCS deprived of growth factor. The cells were plated at a

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concentration of 2 X 10^5 cells/ml in a 96 well microtiter plate (200 μ l/well) with and without growth factor. Recombinant human growth factors were diluted into CHO conditioned medium (CM) before addition to cells. Following incubation for 42 hours. (3 H) thymidine (1 μ Ci/well: New England Nuclear, Boston. MA) was added and the cells were incubated for another 6 hours. The cells were then hypotonically lysed and harvested onto glass fiber filters. The filters were washed with distilled water, dried and counted in liquid scintillation fluid.

Bone Marrow Cultures - Informed consent was obtained prior to 10 aspirating bone marrow from normal volunteers. Aspirated bone marrow was diluted 1:1 in α - medium without nucleosides containing preservative-free sodium heparin. A single cell suspension was prepared, layered, over an equal volume of Ficoll-Hypaque (sp gr 1.077 q/ml) and then centrifuged for 25 minutes at 1,500 rpm at 40°C. The 15 light-density mononuclear cells were collected and washed and diluted to 5 X10⁵ cells/ml with Iscove's modified Dulbecco's medium plus 20% modified FCS (Gibco BRL). Cells (1.25 X 10⁵ /ml) were plated in 0.8% methylcellulose supplemented with various concentrations of rHu Epo, rHu IL-3 and hybrid growth factors. Cultures were incubated for 20 either 7 or 14 days in a humidified atmosphere with 5% ${\rm CO}_2$ at 37°C. Colonies were counted at day 7 for CFU-E and at day 14 for BFU-E under an inverted microscope.

Western Blot Analysis - CHO CM containing approximately 75 ng of IL-3:Epo fusion protein was electrophoresed on a 10-20% gradient SDS PAGE gel (Integrated Separations Systems) under reducing and denaturing conditions. Samples were loaded in 0.0625 M Tris-HCl (pH 6.8), 2% SDS, 5% 2-mercaptoethanol, 10% glycerol and 0.002% bromophenolblue following heat treatment at 100°C for 3 minutes. The proteins were transferred to nitrocellulose (Bio Rad) in 25 mM Tris, 129 mM glycine, pH 8.3, 20% methanol, at 150 V, constant power, for 90 minutes. The transfer efficiency was monitored by visual examination of the completeness of transfer of prestained molecular weight markers (Bio Rad). The nitrocellulose membrane was incubated in PBS containing 3% BSA for 1 hour at room temperature and subsequently washed in PBS

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containing 0.5% Tween (PBS-T) for 5 minutes at room temperature. The membrane was probed with primary anti-Epo anti-sera in 3% BSA in PBS. Excess antibody was removed by 3, 5 minute room temperature washes in PBS-T. The nitrocellulose membrane was then probed with a secondary antibody conjugate (Goat anti-Rabbit IgG/ Alkaline Phosphatase, Bio Rad) for 1 hour at room temperature. Excess secondary antibody was removed by two washes with PBS-T as above. Color development was carried out by incubation with color reagents (Bio Rad) in alkaline phosphatase buffer (100 mM Tris HCl, pH 9.5, 100 mM NaCl, 5 mM MgCl $_2$). The reaction was stopped by immersion of the membrane in cold (4°C) distilled H $_2$ 0.

Results and Discussion

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15 Hybrid growth factor plasmid amplification. Individual transfected CHO cell clones producing significant amounts of the desired hybrid growth factor were identified by ELISA capture assay, Table II. The clones were plated out and placed in medium with increasing concentrations of MSX, ranging between 100 μ M and 500 μ M. Colonies 20 surviving at the highest concentration of MSX were isolated and grown Serum and drug-free medium was then added to the cells and collected after 4 days. At the time of collection fresh serum and drug-free medium was added to the cells. A total of 3 collections were taken. The amount of hybrid growth factor produced in the collections was determined by Epo or G-CSF ELISA assay (Table 25 III) and appropriate collections were pooled. The pooled CM was used as a source of hybrid growth factors in all cellular assays.

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TABLE II

	Hybrid Growth Factor	OD (495 nm)
	CHO CM*	0.24
	IL-3:Epo Flex	2.44
5	IL-3:Epo Short	2.46
	Epo:IL-3 Short	1.59
	IL-3:G-CSF	2.40

* Conditioned medium from CHO cells transfected with the **vector** pEe6.

		TABLE III	
	Hybrid Growth Factor	Collection	Concentration (µg/ml)*
	IL-3:Epo flex	lst	3.0
15		2nd	4.2
		3rd	3.3
	IL-3:Epo Short	lst	1.5
		2nd	5.8
		3rd	6.7
20	Epo:IL-3 Short	1st	26.7
		2nd	53.3
		3rd	58.7
	IL-3:G-CSF	lst	2.2
		2nd	2.0
25		3rd	2.0

* Concentrations were determined by Epo and G-CSF ELISA Assay.

Detection of hybrid growth factor production. In order to confirm that the IL-3 and Epo detected in the ELISA capture assays were being produced in the form of a fusion protein, Western blot analysis was performed. Conditioned medium from CHO cells transfected with IL-3:Epo Flex cDNA (Figure 1, lane 2), IL-3:Epo Short cDNA (Figure 1, lane 3) and Epo:IL-3 Short cDNA (Figure 1, lane 4) were probed with mouse anti-Epo polyclonal anti-sera. Immunoreactive material corresponding to a molecular weight of approximately 50,000 daltons.

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the expected size of the IL-3:Epo and Epo:IL-3 hybrid growth factors, was detected in each sample. Comparison with increasing concentrations of rHu Epo (Figure 1, lanes 6-10) indicated that the antibody used in this analysis recognizes the Epo moiety of the fusion proteins efficiently.

IL-3 bioactivity of the IL-3:Epo and Epo:IL-3 hybrid growth factors. To determine whether the IL-3 moiety of the IL-3:Ēpo and Epo:IL-3 hybrid growth factors was functional, its ability to support growth of the IL-3-dependent human cell line, AML193, was evaluated (Figure 2). As Epo does not support growth of these cells (Figure 2), only IL-3 activity was measured in this assay system. CHO CM containing rHu IL-3 and levels of hybrid growth factors sufficient to support maximal proliferation were added to the culture medium. The cells were then pulsed with (³H) thymidine and the radioactivity incorporated into the DNA was used as a measure of cell growth. Cells exposed to CHO CM containing no growth factors, supported the proliferation of AML193 cells to the same extent as did cells grown in medium alone. Each of the fusion proteins when present in excess, supported the growth of AML193 cells in a manner equivalent to that of rHu IL-3.

The functional activity of the IL-3 portion of the IL-3:Epo and Epo:IL-3 hybrid growth factors was further evaluated by comparing the fusion proteins to rHu IL-3 in dose response experiments (Figure 3). The incorporation of (³H) thymidine into AML193 DNA was again used as a measure of cell proliferation. When IL-3 was located at the N-terminus of the hybrid growth factor protein (IL-3:Epo), its ability to support AML193 proliferation was equivalent to that of rHu IL-3 (ED50 = 5 fmol/ml). Size (2 aa versus 23 aa) and flexibility of the linker did not greatly impact the function of the IL-3 moiety. However, when IL-3 was located at the C-terminus of the fusion protein (Epo:IL-3), its ability to support the growth of AML193 cells was less (ED50 = 200 fmol/ml) than that of rHu IL-3 and the IL-3:Epo hybrid factors. These results suggest that linkage of IL-3 at the N-terminus interferes with function while linkage at the C-terminus does not. It has previously been reported that modification of the C-terminus of

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murine IL-3 did not interfere with its activity (11). Therefore, it should be possible to target any molecule or compound of interest to cells expressing the IL-3 receptor through linkage to IL-3 at its C-terminus.

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Epo bioactivity of the IL-3: Epo and Epo: IL-3 hybrid growth factors. To determine whether the Epo moiety of the hybrid growth factors was functional, its ability to support the growth of the Epo-dependent murine cell line FDC-P1/ER, was evaluated (Figure 4). This line derived from FDC-P1 cells expresses the murine Epo receptor (10), and is dependent on either murine IL-3 or Epo (murine and human) for growth (Figure 4). As IL-3 is a species specific growth factor, murine IL-3-dependent cells do not respond to human IL-3 (12). Therefore, when using the FDC-P1/ER cell line to evaluate functionality, only the activity of the Epo moiety is measured. CHO CM containing rHu Epo and levels of hybrid growth factors sufficient to support maximal proliferation were added to the culture medium. The cells were then pulsed with (3H) thymidine and the radioactivity incorporated into the DNA was used as a measure of cell growth. Cells exposed to CHO CM which did not contain cytokines did not support the proliferation of FDC-P1/ER cells. Each of the fusion proteins when present in excess, stimulated the growth of FDC-P1/ER cells to the same extent as did rHu Epo. (Figure 4)

The biological function of the Epo portion of the IL-3:Epo and Epo:IL-3 hybrid growth factors was further analyzed by comparing the fusion proteins to rHu Epo in dose response experiments (Figure 5). The incorporation of (³H) thymidine into FDC-P1/ER cells was used as a measure of cell proliferation. Each of the hybrid growth factors was equivalent to rHu Epo in ability to stimulate proliferation of FDC-P1/ER cells (ED50 = 50 fmol/ml). Size (2-3 aa versus 23 aa) and flexibility of the linker, as well as the orientation of Epo within the protein (N-terminus versus C-terminus) did not alter function. Evidence exists suggesting that the N-terminus of Epo is not involved in receptor binding as a monoclonal antibody directed toward the N-terminus of Epo does not neutralize its activity (13). The results

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presented here suggest that linkage of Epo to a second protein does not impair its ability to bind its receptor or transduce a signal. Epo could therefore be useful as a carrier protein which would target a molecule or compound of interest to those cells expressing the Epo receptor.

IL-3 plus Epo bioactivity of the IL-3:Epo and Epo:IL-3 hybrid growth factors. In order to study the effects of IL-3 and Epo in combination, proliferation of a human cell line, TF-1 (7), dependent on IL-3 and Epo for growth was measured. This experiment was done on a cytokine weight basis and the results are represented on a molar basis (Figure 6). rHu IL-3 (R & D Systems) made in E. coli is nonglycosylated. rHu Epo and hybrid growth factors made in CHO cells are glycosylated. Therefore, when equal weights of the growth factors were added to the cell culture medium, approximately twice the number of unglycosylated molecules of IL-3 were added as compared to glycosylated Epo and hybrid growth factor molecules.

CHO CM containing rhu IL-3 plus rhu Epo and levels of hybrid growth factors which support suboptimal proliferation of TF-1 cells adapted for growth in IL-3 were added to the culture medium. Cell growth was monitored by radioactivity incorporated into the DNA. (Figure 6) The activities of IL-3 plus Epo were not synergistic in this cell line, nor were they additive. At these low levels, the activities of the IL-3:Epo Flex and IL-3:Epo Short fusion proteins were comparable to those of a mixture of the two cytokines. Epo:IL-3 Short activity was again reduced in comparison to that of the IL-3:Epo hybrid growth factors and the combination of IL-3 plus Epo. This is likely to be due to decreased IL-3 activity.

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The biological activity of the IL-3:Epo and Epo:IL-3 hybrid growth factors was further evaluated in dose response experiments (Figures 7 & 8). TF-1 cells adapted for optimal growth in IL-3 were exposed to CHO CM containing hybrid growth factors (Figure 7). Each of the fusion proteins when present in excess were able to support growth of the cells to the same extent (Figure 7A). At lower doses, the IL-

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3:Epo Flex protein appeared to be slightly more potent than the short linkered hybrids (IL-3:Epo Flex ED50 = 0.37 fmol/ml; IL-3:Epo Short ED50 = 0.75 fmol/ml; Epo:IL-3 Short ED50 = 0.9 fmol/ml) (Figure 7B). This result was more pronounced in experiments done with TF-1 cells adapted for optimal growth in GM-CSF (Figure 8). The IL-3:Epo Flex protein was dramatically more potent than the hybrid growth factors containing short linkers (IL-3:Epo Flex ED50 = 0.07 fmol/ml; IL-3:Epo Short and Epo:IL-3 Short ED50 = 0.75 fmol/ml (Figure 8B). When present in excess with the GM-CSF adapted TF-1 cells, each of the fusion proteins stimulated cell proliferation to a similar extent (Figure 8A). These results suggest that when IL-3 and Epo are fused, the 23 aa flexible linker allows more efficient receptor interaction than does a short (2-3 aa) linker.

It appears that induction of receptor expression is possible by growing a cell in the presence of a cytokine whose receptor it has the potential to express. An up regulation of Epo receptor expression has been reported in IL-3-dependent cells transferred into medium Thus, it is likely that growing TF-1 supplemented with Epo (14). cells in IL-3 or GM-CSF, preferentially increases the appearance of cell surface IL-3 or GM-CSF receptors. Several research groups (15-20) have observed a subset population of GM-CSF and IL-3 receptors on primary human cells and hematopoietic cell lines capable of binding both GM-CSF and IL-3. It has been suggested that a single accessory molecule preferentially interacts with this subset of GM-CSF/IL-3 receptors allowing the transduction of signal. Our results raise the possibility that GM-CSF could be inducing the expression of an accessory molecule in TF-1 cells which may be important for binding and could possibly link IL-3:Epo signal transduction. This protein could be identical to the GM-CSF/IL-3 receptor accessory protein.

Erythroid colony formation stimulated by IL-3:Epo and Epo:IL-3 hybrid growth factors. To assess the biological activity of the fusion proteins on normal hematopoietic progenitor cells, analysis of the formation of erythroid (BFU-E and CFU-E) colonies from nonadherent mononuclear human bone marrow cells was performed. (Table IV) As was

observed with the cell lines, the Epo moiety of each of the fusion proteins was equally active (CFU-E formation) on bone marrow progenitor cells. The IL-3:Epo Flex protein was the most active hybrid growth factor while the Epo:IL-3 Short protein was the least. (It stimulated two-thirds the number of BFU-E as did the IL-3:Epo Flex.) These results suggest that the IL-3:Epo Flex fusion protein may have significant clinical benefits where indications for combination therapy with IL-3 and Epo may prove efficacious.

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		TABLE IV	
	Growth Factors	<u>BFU-E</u> a,b	<u>CFU-E</u> C
	No Factors	-	-
	0.2 pmol/ml IL-3	-	-
15	0.1 pmol/ml Epo	+	+++
	0.2 pmol/ml IL-3 &		
0.1 pmol/ml Epo 0.1 pmol/ml IL-3:Epo Flex	0.1 pmol/ml Epo	+++	+++
	+++	+++	
	0.1 pmol/ml IL-3:Epo Short	++	+++
20	0.1 pmol/ml Epo:IL-3 Short	+	+++

- a Mononuclear human bone marrow cells were used as a target cell population.
- b BFU-E were counted 14 days after plating.
- 25 c CFU-E were counted 7 days after plating.

IL-3 bioactivity of the IL-3:G-CSF hybrid growth factor. To determine whether the IL-3 moiety of the IL-3:G-CSF hybrid growth factor was functional, its ability to support growth of the IL-3-dependent human cell line TF-1, was evaluated in a dose response experiment (Figure 9). Quantitation of IL-3:G-CSF protein in CHO CM was performed using a G-CSF ELISA assay in which the standard is unglycosylated G-CSF. Since the IL-3:G-CSF fusion protein is glycosylated, measurements are approximate. G-CSF does not support growth of TF-1 cells (Figure 9), therefore, the only activity measured in this assay system was IL-3.

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CHO CM containing rhu IL-3, rHu G-CSF, and IL-3:G-CSF hybrid growth factor were added to the culture medium. The radioactivity incorporated into the DNA was used as a measure of cell proliferation. CHO CM did not support growth of TF-1 cells. The mixture of rhu IL-3 plus rHu G-CSF stimulated proliferation to the same extent as did rhu IL-3 alone. The IL-3:G-CSF hybrid growth factor induced a dose response similar to that observed with IL-3.

G-CSF bioactivity of the IL-3:G-CSF hybrid growth factor. To evaluate the biological function of the G-CSF moiety of the IL-3:G-CSF hybrid growth factor, its ability to stimulate proliferation of the murine cell line, NSF-60, was tested. (Figure 10) G-CSF, unlike IL-3 is not species specific, therefore, human G-CSF will actively support growth of murine cells (21). Cells exposed to CHO CM containing no growth factors, supported the proliferation of NSF-60 cells to the same extent as did cells grown in medium alone. The IL-3:G-CSF hybrid growth factor stimulated growth in a dose dependent manner equivalent to that observed with G-CSF.

IL-3 plus G-CSF bioactivity of the IL-3:G-CSF hybrid growth factor. The biological function of the IL-3:G-CSF hybrid growth factor was evaluated by its ability to support growth of an IL-3-, G-CSF-dependent human cell line, AML193. CHO CM containing rHu IL-3, rHu G-CSF and IL-3:G-CSF hybrid growth factor were added to the culture medium. Cell proliferation was monitored by incorporation of radioactivity into the DNA. (Figure 11). Both IL-3 and G-CSF supported growth of this cell line in a dose dependent manner. The two cytokine activities were not synergistic, nor were they additive. The IL-3:G-CSF hybrid growth factor stimulated AML193 proliferation to a greater extent than did the mixture of the two cytokines.

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- 30 -

SEQUENCE LISTING

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 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
- (vi) CURRENT APPLICATION DATA:
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 - (C) TELEX: 844-481
- (2) INFORMATION FOR SEQ ID NO:1:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 133 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein

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- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

Ala Pro Met Thr Gln Thr Thr Ser Leu Lys Thr Ser Trp Val Asn Cys
1 5 10 15

Ser Asn Met Ile Aso Glu Ile Ile Thr His Leu Lys Gln Pro Pro Leu 20 25 30

Pro Leu Leu Asp Phe Asn Asn Leu Asn Gly Glu Asp Gln Asp Ile Leu 35 40 45

Met Glu Asn Asn Leu Arg Arg Pro Asn Leu Glu Ala Phe Asn Arg Ala 50 55 60

Val Lys Ser Leu Gln Asn Ala Ser Ala Ile Glu Ser Ile Leu Lys Asn 65 70 75 80

Leu Leu Pro Cys Leu Pro Leu Ala Thr Ala Ala Pro Thr Arg His Pro 85 90 95

Ile His Ile Lys Asp Gly Asp Trp Asn Glu Phe Arg Arg Lys Leu Thr

Phe Tyr Leu Lys Thr Leu Glu Asn Ala Gln Ala Gln Gln Thr Thr Leu 115 120 125

Ser Leu Ala Ile Phe 130

(2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 79 amino acids
 - (R) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (v) FRAGMENT TYPE: N-terminal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Ala Pro Met Thr Gln Thr Thr Ser Leu Lys Thr Ser Trp Val Asn Cys
1 10 15

Ser Asn Met Ile Asp Glu Ile Ile Thr His Leu Lys Gln Pro Pro Leu 20 25 30

Pro Leu Leu Asp Phe Asn Asn Leu Asn Gly Glu Asp Gln Asp Ile Leu 35 40 45

Met Glu Asn Asn Leu Arg Arg Pro Asn Leu Glu Ala Phe Asn Arg Ala 50 55 60

Val Lys Ser Leu Gln Asn Ala Ser Ala Ile Glu Ser Ile Leu Lys 65 70 75

(2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 166 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

Ala Pro Pro Arg Leu Ile Cys Asp Ser Arg Val Leu Glu Arg Tyr Leu 1 5 10 15

Leu Glu Ala Lys Glu Ala Glu Asn Ile Thr Thr Gly Cys Ala Glu His 20 25 30

Cys Ser Leu Asn Glu Asn Ile Thr Val Pro Asp Thr Lys Val Asn Phe 35 40 45

Tyr Ala Trp Lys Arg Met Glu Val Gly Gln Gln Ala Val Glu Val Trp 50 55 60

Gln Gly Leu Ala Leu Leu Ser Glu Ala Val Leu Arg Gly Gln Ala Leu 65 70 75 80

Leu Val Asn Ser Ser Gln Pro Trp Glu Pro Leu Gln Leu His Val Asp 85 90 95

Lys Ala Val Ser Gly Leu Arg Ser Leu Thr Thr Leu Leu Arg Ala Leu 100 105 110 Gly Ala Gln Lys Glu Ala Ile Ser Pro Pro Asp Ala Ala Ser Ala Ala 115 120 125

Pro Leu Arg Thr Ile Thr Ala Asp Thr Phe Arg Lys Leu Phe Arg Val 130 135 140

Tyr Ser Asn Phe Leu Arg Gly Lys Leu Lys Leu Tyr Thr Gly Glu Ala 145 150 155 160

Cys Arg Thr Gly Asp Arg 165

(2) INFORMATION FOR SEQ ID NO:4:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 155 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Cys Asp Ser Arg Val Leu Glu Arg Tyr Leu Leu Glu Ala Lys Glu Ala 10 15

Glu Asn Ile Thr Thr Gly Cys Ala Glu His Cys Ser Leu Asn Glu Asn 20 25 30

Ile Thr Val Pro Asp Thr Lys Val Asn Phe Tyr Ala Trp Lys Arg Met 45

Glu Val Gly Gln Gln Ala Val Glu Val Trp Gln Gly Leu Ala Leu Leu 50 55 60

Ser Glu Ala Val Leu Arg Gly Gln Ala Leu Leu Val Asn Ser Ser Gln 80

Pro Trp Glu Pro Leu Gln Leu His Val Asp Lys Ala Val Ser Gly Leu 85 90 95

Arg Ser Leu Thr Thr Leu Leu Arg Ala Leu Gly Ala Gln Lys Glu Ala 100 105 110 Ile Ser Pro Pro Asp Ala Ala Ser Ala Ala Pro Leu Arg Thr Ile Thr 120

Ala Asp Thr Phe Arg Lys Leu Phe Arg Val Tyr Ser Asn Phe Leu Arg

Gly Lys Leu Lys Leu Tyr Thr Gly Glu Ala Cys

(2) INFORMATION FOR SEQ ID NO:5:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 174 amino acids

 - (B) TYPE: amino acid
 (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

Thr Pro Leu Gly Pro Ala Ser Ser Leu Pro Gln Ser Phe Leu Leu Lys

Cys Leu Glu Gln Val Arg Lys Ile Gln Gly Asp Gly Ala Ala Leu Gln

Glu Lys Leu Cys Ala Thr Tyr Lys Leu Cys His Pro Glu Glu Leu Val 35 40 45

Leu Leu Gly His Ser Leu Gly Ile Pro Trp Ala Pro Leu Ser Ser Cys

Pro Ser Gln Ala Leu Gln Leu Ala Gly Cys Leu Ser Gln Leu His Ser

Gly Leu Phe Leu Tyr Gln Gly Leu Leu Gln Ala Leu Glu Gly Ile Ser

Pro Glu Leu Gly Pro Thr Leu Asp Thr Leu Gln Leu Asp Val Ala Asp

Phe Ala Thr Thr Ile Trp Gln Gln Met Glu Glu Leu Gly Met Ala Pro

Ala Leu Gln Pro Thr Gln Gly Ala Met Pro Ala Phe Ala Ser Ala Phe 135

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Gln Arg Arg Ala Gly Gly Val Leu Val Ala Ser His Leu Gln Ser Phe 145 150 155 160

Leu Glu Val Ser Tyr Arg Val Leu Arg His Leu Ala Gln Pro 165 170

(2) INFORMATION FOR SEQ ID NO:6:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 302 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Ala Pro Met Thr Gln Thr Thr Ser Leu Lys Thr Ser Trp Val Asn Cys
10 15

Ser Asn Met Ile Asp Glu Ile Ile Thr His Leu Lys Gln Pro Pro Leu 20 25 30

Pro Leu Leu Asp Phe Asn Asn Leu Asn Gly Glu Asp Gln Asp Ile Leu 35 40 45

Met Glu Asn Asn Leu Arg Arg Pro Asn Leu Glu Ala Phe Asn Arg Ala 50 55

Val Lys Ser Leu Gln Asn Ala Ser Ala Ile Glu Ser Ile Leu Lys Asn 65 70 75 80

Leu Leu Pro Cys Leu Pro Leu Ala Thr Ala Ala Pro Thr Arg His Pro

Ile His Ile Lys Asp Gly Asp Trp Asn Glu Phe Arg Arg Lys Leu Thr

Phe Tyr Leu Lys Thr Leu Glu Asn Ala Gln Ala Gln Gln Thr Thr Leu 115 120 125

Ser Leu Ala Ile Phe Leu Asp Met Ala Pro Pro Arg Leu Ile Cys Asp 130 135 140

Ser Arg Val Leu Glu Arg Tyr Leu Leu Glu Ala Lys Glu Ala Glu Asn 145 150 155 160 Ile Thr Thr Gly Cys Ala Glu His Cys Ser Leu Asn Glu Asn Ile Thr 165 170 175

Val Pro Asp Thr Lys Val Asn Phe Tyr Ala Trp Lys Arg Met Glu Val 180 185 190

Gly Gln Gln Ala Val Glu Val Trp Gln Gly Leu Ala Leu Leu Ser Glu 195 200 205

Ala Val Leu Arg Gly Gln Ala Leu Leu Val Asn Ser Ser Gln Pro Trp 210 215 220

Glu Pro Leu Gln Leu His Val Asp Lys Ala Val Ser Gly Leu Arg Ser 225 230 235 240

Leu Thr Thr Leu Leu Arg Ala Leu Gly Ala Gln Lys Glu Ala Ile Ser 245 250 255

Pro Pro Asp Ala Ala Ser Ala Ala Pro Leu Arg Thr Ile Thr Ala Asp 260 265 270

Thr Phe Arg Lys Leu Phe Arg Val Tyr Ser Asn Phe Leu Arg Gly Lys 275 280 285

Leu Lys Leu Tyr Thr Gly Glu Ala Cys Arg Thr Gly Asp Arg 290 295 300

(2) INFORMATION FOR SEQ ID NO:7:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 321 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

Ala Pro Met Thr Gln Thr Thr Ser Leu Lys Thr Ser Trp Val Asn Cys
1 10 15

Ser Asn Met Ile Asp Glu Ile Ile Thr His Leu Lys Gln Pro Pro Leu 20 25 30

Pro Leu Leu Asp Phe Asn Asn Leu Asn Gly Glu Asp Gln Asp Ile Leu 35 40 45

Met	G lu 50	Asn	Asn	Leu	Arg	Arg 55	Pro	Asn	Leu	Glu	A1a 60	Phe	Asn	Arg	Ala
Va 1 65	Lys	Ser	Leu	Gln	Asn 70	Ala	Ser	Ala	Ile	G 1 u 75	Ser	Ile	Leu	Lys	Asn 80
Leu	Leu	Pro	Cys	Leu 85	Pro	Leu	Ala	Thr	Ala 90	Ala	Pro	Thr	Arg	His 95	Pro
Пe	His	Ile	Lys 100	Asp	Gly	Asp	Trp	Asn 105	Glu	Phe	Arg	Arg	Lys 110	Leu	Thr
Phe	Tyr	Leu 115	Lys	Thr	Leu	Glu	Asn 120	Ala	Gln	Ala	Gln	Gln 125	Thr	Thr	Leu
Ser	Leu 130	Ala	Ser	Gly	Gly	Gly 135	Gly	Ser	Gly	Gly	Gly 140	Gly	Ser	Gly	Gly
Gly 145	Gly	Ser	Ala	Leu	Ala 150	Ile	Phe	Leu	Asp	Met 155	Ala	Pro	Pro	Arg	Leu 160
Пe	Cys	Asp	Ser	Arg 165	Val	Leu	G1u	Arg	Tyr 170	Leu	Leu	Glu	Ala	Lys 175	Glu
Ala	6 lu	Asn	Ile 180	Thr	Thr	Gly	Cys	Ala 185	Glu	His	Cys	Ser	Leu 190	Asn	Glu
Asn	I le	Thr 195	Va 1	Pro	Asp	Thr	Lys 200	Val	Asn	Phe	Tyr	A1a 205	Trp	Lys	Arg
Met	G 1u 210	Val	Gly	Gln	Gln	Ala 215	Va 1	Glu	Val	Trp	G1n 220	Gly	Leu	Ala	Leu
Leu 225	Ser	Glu	Ala	Va 1	Leu 230	Arg	Gly	G1n	Ala	Leu 235	Leu	Val	Asn	Ser	Ser 240
Gln	Pro	Trp	G lu	Pro 245	Leu	Gln	Leu	His	Va 1 250	Asp	Lys	Ala	Val	Ser 255	Gly
Leu	Arg	Ser	Leu 260	Thr	Thr	Leu	Leu	Arg 265	Ala	Leu	Gly	Ala	G1n 270	Lys	Glu
Ala	I le	Ser 275	Pro	Pro	Asp	Ala	A1a 280	Ser	Ala	Ala	Pro	Leu 285	Arg	Thr	Ile
Ţhr	A 1a 290	Asp	Thr	Phe	Arg	Lys 295	Leu	Phe	Arg	Val	Tyr 300	Ser	Asn	Phe	Leu
Arg 305	Gly	Lys	Leu	Lys	Leu 310	Tyr	Thr	Gly	G1u	A1a 315	Cys	Arg	Thr	Gly	Asp 320

Arg

(2) INFORMATION FOR SEQ ID NO:8:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 303 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

Ala Pro Pro Arg Leu Ile Cys Asp Ser Arg Val Leu Glu Arg Tyr Leu 1 10 15

Leu Glu-Ala Lys Glu Ala Glu Asn Ile Thr Thr Gly Cys Ala Glu His 20 25 30

Cys Ser Leu Asn Glu Asn Ile Thr Val Pro Asp Thr Lys Val Asn Phe 35 40 45

Tyr Ala Trp Lys Arg Met Glu Val Gly Gln Gln Ala Val Glu Val Trp
50 55 60

Gln Gly Leu Ala Leu Leu Ser Glu Ala Val Leu Arg Gly Gln Ala Leu 65 70 75 80

Leu Val Asn Ser Ser Gln Pro Trp Glu Pro Leu Gln Leu His Val Asp 85 90 95

Lys Ala Val Ser Gly Leu Arg Ser Leu Thr Thr Leu Leu Arg Ala Leu 100 105 110

Gly Ala Gln Lys Glu Ala Ile Ser Pro Pro Asp Ala Ala Ser Ala Ala 115 120 125

Pro Leu Arg Thr Ile Thr Ala Asp Thr Phe Arg Lys Leu Phe Arg Val 130 135 140

Tyr Ser Asn Phe Leu Arg Gly Lys Leu Lys Leu Tyr Thr Gly Glu Ala 145 150 155 160

Cys Arg Thr Gly Asp Arg Ala Ala Ala Met Ala Pro Met Thr Gln Thr 165 170 175

Thr Ser Leu Lys Thr Ser Trp Val Asn Cys Ser Asn Met Ile Asp Glu 180 185 190 Ile Ile Thr His Leu Lys Gln Pro Pro Leu Pro Leu Leu Asp Phe Asn 195 200 205.

Asn Leu Asn Gly Glu Asp Gln Asp Ile Leu Met Glu Asn Asn Leu Arg 210 215 220

Arg Pro Asn Leu Glu Ala Phe Asn Arg Ala Val Lys Ser Leu Gln Asn 225 230 235 240

Ala Ser Ala Ile Glu Ser Ile Leu Lys Asn Leu Leu Pro Cys Leu Pro 245 250 255

Leu Ala Thr Ala Ala Pro Thr Arg His Pro Ile His Ile Lys Asp Gly 260 265 270

Asp Trp Asn Glu Phe Arg Arg Lys Leu Thr Phe Tyr Leu Lys Thr Leu 275 280 285

Glu Asn Ala Gln Ala Gln Gln Thr Thr Leu Ser Leu Ala Ile Phe 290 295 300

(2) INFORMATION FOR SEQ ID NO:9:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 322 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

Ala Pro Pro Arg Leu Ile Cys Asp Ser Arg Val Leu Glu Arg Tyr Leu 1 10 15

Leu Glu Ala Lys Glu Ala Glu Asn Ile Thr Thr Gly Cys Ala Glu His

Cys Ser Leu Asn Glu Asn Ile Thr Val Pro Asp Thr Lys Val Asn Phe

Tyr Ala Trp Lys Arg Met Glu Val Gly Gln Gln Ala Val Glu Val Trp

Gln Gly Leu Ala Leu Leu Ser Glu Ala Val Leu Arg Gly Gln Ala Leu 65 70 75 80 Leu Val Asn Ser Ser Gln Pro Trp Glu Pro Leu Gln Leu His Val Asp 85 Lys Ala Val Ser Gly Leu Arg Ser Leu Thr Thr Leu Leu Arg Ala Leu 105 Gly Ala Gln Lys Glu Ala Ile Ser Pro Pro Asp Ala Ala Ser Ala Ala 125 Pro Leu Arg Thr Ile Thr Ala Asp Thr Phe Arg Lys Leu Phe Arg Val Tyr Ser Asn Phe Leu Arg Gly Lys Leu Lys Leu Tyr Thr Gly Glu Ala 150 155 Cys Arg Thr Gly Asp Arg Ala Ala Ala Ser Gly Gly Gly Ser Gly Gly Gly Gly Ser Gly Gly Gly Ser Ala Ala Ala Met Ala Pro Met 180 185 190 Thr Gln Thr Thr Ser Leu Lys Thr Ser Trp Val Asn Cys Ser Asn Met 200 Ile Asp Glu Ile Ile Thr His Leu Lys Gln Pro Pro Leu Pro Leu Leu 210 220 Asp Phe Asn Asn Leu Asn Gly Glu Asp Gln Asp Ile Leu Met Glu Asn Asn Leu Arg Arg Pro Asn Leu Glu Ala Phe Asn Arg Ala Val Lys Ser 255 245 250 Leu Gln Asn Ala Ser Ala Ile Glu Ser Ile Leu Lys Asn Leu Leu Pro 265

Cys Leu Pro Leu Ala Thr Ala Ala Pro Thr Arg His Pro Ile His Ile

Lys Asp Gly Asp Trp Asn Glu Phe Arg Arg Lys Leu Thr Phe Tyr Leu

Lys Thr Leu Glu Asn Ala Gln Ala Gln Gln Thr Thr Leu Ser Leu Ala

285

315

280

Ile Phe

305

(2) INFORMATION FOR SEQ ID NO:10:

275

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 317 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single

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(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

Ala Pro Met Thr Gln Tnr Thr Ser Leu Lys Thr Ser Trp Val Asn Cys
1 10 15

Ser Asn Met Ile Asp Glu Ile Ile Thr His Leu Lys Gln Pro Pro Leu 20 25 30

Pro Leu Leu Asp Phe Asn Asn Leu Asn Gly Glu Asp Gln Asp Ile Leu 35 40 45

Met Glu Asn Asn Leu Arg Arg Pro Asn Leu Glu Ala Phe Asn Arg Ala 50 55 60

Val Lys Ser Leu Gln Asn Ala Ser Ala Ile Glu Ser Ile Leu Lys Asn 65 70 75 80

Leu Leu Pro Cys Leu Pro Leu Ala Thr Ala Ala Pro Thr Arg His Pro 85 90 95

Ile His Ile Lys Asp Gly Asp Trp Asn Glu Phe Arg Arg Lys Leu Thr

Phe Tyr Leu Lys Thr Leu Glu Asn Ala Gln Ala Gln Gln Thr Thr Leu 115 120 125

Ser Leu Ala Ile Phe Leu-Glu Ala Ala Ser Leu Pro Ala Met Thr 130 135 140

Pro Leu Gly Pro Ala Ser Ser Leu Pro Gln Ser Phe Leu Leu Lys Cys 145 150 155 160

Leu Glu Gln Val Arg Lys Ile Gln Gly Asp Gly Ala Ala Leu Gln Glu 165 170 175

Lys Leu Cys Ala Thr Tyr Lys Leu Cys His Pro Glu Glu Leu Val Leu 180 185 190

Leu Gly His Ser Leu Gly Ile Pro Trp Ala Pro Leu Ser Ser Cys Pro 195 200 205

Ser Gln Ala Leu Gln Leu Ala Gly Cys Leu Ser Gln Leu His Ser Gly 210 215 220

Leu Phe Leu Tyr Gln Gly Leu Leu Gln Ala Leu Glu Gly Ile Ser Pro 225 230 235 240

Glu Leu Gly Pro Thr Leu Asp Thr Leu Gln Leu Asp Val Ala Asp Phe 245 250 255

Ala Thr Thr Ile Trp Gln Gln Met Glu Glu Leu Gly Met Ala Pro Ala 260 265 270

Leu Gln Pro Thr Gln Gly Ala Met Pro Ala Phe Ala Ser Ala Phe Gln 275 280 285

Arg Arg Ala Gly Gly Val Leu Val Ala Ser His Leu Gln Ser Phe Leu 290 295 300

Glu Val Ser Tyr Arg Val Leu Arg His Leu Ala Gln Pro . 305 310 315

(2) INFORMATION FOR SEQ ID NO:11:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 994 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: both
- (ii) MOLECULE TYPE: DNA (genomic)
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (ix) FEATURE:
 - (A) NAME/KEY: CDS (B) LOCATION: 14..977
- (ix) FEATURE:
 - (A) NAME/KEY: mat_peptide
 (B) LOCATION: 71..977
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:
- AATTGCCGCC ACC ATG AGC CGC CTG CCC GTC CTG CTC CTG CTC CAA CTC

 Met Ser Arg Leu Pro Val Leu Leu Leu Gln Leu

 -19

 -15

 -10
- CTG GTC CGC CCC GGA CTC CAA GCT CCC ATG ACC CAG ACA ACT AGT TTG

 Pro Gly Leu Gln Ala Pro Met Thr Gln Thr Thr Ser Leu

 -5

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 5

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											GAT Asp					145
											TTC Phe					193
GGG Gly	GAA G lu	GAC Asp	CAA G1n 45	GAC Asp	ATT Ile	CTG Leu	ATG Met	GAA Glu 50	AAT Asn	AAC Asn	CTT Leu	CGA Arg	AGG Arg 55	CCA Pro	AAC Asn	241
CTG Leu	GAG Glu	GCA Ala 60	TTC Phe	AAC Asn	AGG Arg	GCT Ala	GTC Val 65	AAG Lys	AGT Ser	TTA Leu	CAG Gln	AAT Asn 70	GCA Ala	TCA Ser	GCA Ala	289
ATT Ile	GAG Glu 75	AGC Ser	ATT Ile	CTT Leu	AAA Lys	AAT Asn 80	CTC Leu	CTG Leu	CCA Pro	TGT Cys	CTG Leu 85	CCC Pro	CTG Leu	GCC Ala	ACG Thr	337
GCC Ala 90	GCA Ala	CCC Pro	ACG Thr	CGA Arg	CAT His 95	CCA Pro	ATC Ile	CAT His	ATC Ile	AAG Lys 100	GAC Asp	GGT Gly	GAC Asp	TGG Trp	AAT Asn 105	385
GAA Glu	TTC Phe	CGG Arg	AGG Arg	AAA Lys 110	CTG Leu	ACG Thr	TTC Phe	TAT Tyr	CTG Leu 115	AAA Lys	ACC Thr	CTT Leu	GAG G lu	AAT Asn 120	GCG Ala	433
CAG Gln	GCT Ala	CAA Gln	CAG G]n 125	ACG Thr	ACT Thr	TTG Leu	TCG Ser	CTA Leu 130	GCG Ala	ATC Ile	TTT Phe	CTA Leu	GAC Asp 135	ATG Met	GCC Ala	481
CCA Pro	CCA Pro	CGC Arg 140	CTC Leu	ATC Ile	TGT Cys	Asp	AGC Ser 145	CGA Arg	GTC Val	CTG Leu	GAG Glu	AGG Arg 150	TAC Tyr	CTC Leu	TTG Leu	529
GAG Glu	GCC Ala 155	AAG Lys	GAG Glu	GCC Ala	GAG G lu	AAT Asn 160	ATC Ile	ACG Thr	ACG Thr	GGC Gly	TGT Cys 165	GCT Ala	GAA Glu	CAC His	TGC Cys	577
AGC Ser 170	TTG Leu	AAT Asn	GAG G lu	AAT Asn	ATC Ile 175	ACT Thr	GTC Val	CCA Pro	GAC Asp	ACC Thr 180	AAA Lys	GTT Val	AAT Asn	TTC Phe	TAC Tyr 185	6 25
GCG Ala	TGG Trp	AAG Lys	AGG Arg	ATG Met 190	GAG Glu	GTC Val	GGC Gly	CAG Gln	CAG G ln 195	GCC Ala	GTA Val	GAA G lu	GTC Val	TGG Trp 200	CAG Gln	673
GGC Gly	CTG Leu	GCC Ala	CTG Leu 205	CTG Leu	TCG Ser	GAA G lu	GCT Ala	GTC Val 210	CTG Leu	CGG Arg	GGC Gly	CAG Gln	GCC Ala 215	CTG Leu	TTG Leu	721

-	44	-
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								CTG Leu			,	769
								CTT Leu 245			8	817
								GCC Ala			8	865
								CTC Leu			(913
								ACA Thr			9	961
	GAC Asp	T GA	ATAAG	GATO	C CGA	VATT (2				9	994

(2) INFORMATION FOR SEQ ID NO:12:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1015 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: both
- (ii) MOLECULE TYPE: DNA (genomic)
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (ix) FEATURE:
 - (A) NAME/KEY: CDS (B) LOCATION: 8..998
- (ix) FEATURE:
 - (A) NAME/KEY: mat_peptide
 - (B) LOCATION: 89..998

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

TCGAGCC ATG GGG GTG CAC GAA TGT CCT GCC TGG CTG TGG CTT CTC CTG 49 Met Gly Val His Glu Cys Pro Ala Trp Leu Trp Leu Leu Leu -27 -25 -20

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TCC Ser	CTG Leu	CTG Leu	TCG Ser -10	CTC Leu	CCT Pro	CTG Leu	GGC Gly	CTC Leu -5	CCA Pro	GTC Val	CTG Leu	GGC Gly	GCC Ala l	CCA Pro	CCA Pro	97
CGC Arg	CTC Leu 5	ATC Ile	TGT Cys	GAC Asp	AGC Ser	CGA Arg 10	GTC Val	CTG Leu	GAG G1u	AGG Arg	TAC Tyr 15	CTC Leu	TTG Leu	GAG Glu	GCC Ala	145
AAG Lys 20	GAG Glu	GCC Ala	GAG G lu	AAT Asn	ATC Ile 25	ACG Thr	ACG Thr	GGC Gly	TGT Cys	GCT Ala 30	GAA Glu	CAC His	TGC Cys	AGC Ser	TTG Leu 35	193
AAT Asn	GAG G lu	AAT Asn	ATC Ile	ACT Thr 40	GTC Val	CCA Pro	GAC Asp	ACC Thr	AAA Lys 45	GTT Val	AAT Asn	TTC Phe	TAC Tyr	GCG Ala 50	TGG Trp	241
AAG Lys	AGG Arg	ATG Met	GAG Glu 55	GTC Val	GGC Gly	CAG Gìn	CAG Gln	GCC Ala 60	GTA Val	GAA Glu	GTC Val	TGG Trp	CAG Gln 65	GGC Gly	CTG Leu	289
GCC Ala	CTG Leu	CTG Leu 70	TCG Ser	GAA Glu	GCT Ala	GTC Val	CTG Leu 75	CGG Arg	GGC Gly	CAG Gln	GCC Ala	CTG Leu 80	TTG Leu	GTC Va 1	AAC Asn	337
TCG Ser	AGC Ser 85	CAG Gln	CCG Pro	TGG Trp	GAG G lu	CCC Pro 90	CTG Leu	CAA Gln	CTG Leu	CAT His	GTG Val 95	GAT Asp	AAA Lys	GCC Ala	GTC Val	385
AGT Ser 100	GGC Gly	CTT Leu	CGC Arg	AGC Ser	CTC Leu 105	ACC Thr	ACT Thr	CTG Leu	CTT Leu	CGG Arg 110	GCT Ala	CTG Leu	GGA Gly	GCT Ala	CAG Gln 115	433
AAG Lys	GAA G lu	GCC Ala	ATC Ile	TCC Ser 120	CCT Pro	CCA Pro	GAT Asp	GCG Ala	GCC Ala 125	TCA Ser	GCT Ala	GCT Ala	CCA Pro	CTC Leu 130	CGA Arg	481
ACA Thr	ATC Ile	ACT Thr	GCT Ala 135	GAC Asp	ACT Thr	TTC Phe	CGC Arg	AAA Lys 140	CTC Leu	TTC Phe	CGA Arg	GTC Val	TAC Tyr 145	TCC Ser	AAT Asn	529
TTC Phe	CTC Leu	CGG Arg 150	GGA Gly	AAG Lys	CTG Leu	AAG Lys	CTG Leu 155	TAC Tyr	ACA Thr	GGG Gly	GAG G 1 u	GCA Ala 160	TGC Cys	ÄGG Arg	ACA Thr	577
GGG Gly	GAC Asp 165	AGA Arg	GCG Ala	GCC Ala	GCC Ala	ATG Met 170	GCT Ala	CCC Pro	ATG Met	ACC Thr	CAG G 1n 175	ACA Thr	ACT Thr	AGT Ser	TTG Leu	625
AAG Lys 180	ACA Thr	AGC Ser	TGG Trp	GTT Val	AAC Asn 185	TGC Cys	TCT Ser	AAC Asn	ATG Met	ATC Ile 190	GAT Asp	GAA G lu	ATT Ile	ATA I le	ACA Thr 195	673

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	TTA Leu										721
	GAA G lu										769
	GAG G lu										817
	GAG Glu 245										865
	GCA Ala										913
	TTC Phe										961
	GCT Ala						T A	STAA	GGAT(3	1008
CGA	ATTC										1015

(2) INFORMATION FOR SEQ ID NO:13:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1039 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: both
- (ii) MOLECULE TYPE: DNA (genomic)
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (ix) FEATURE:

 - (A) NAME/KEY: CDS
 (B) LOCATION: 14..1021
- (ix) FEATURE:
 - (A) NAME/KEY: mat_peptide (B) LOCATION: 71..1021

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

	(^	.,	-400,			• • • •	•	4	•••		•			
AA	TTGC(CGCC	ACC								CTC Leu			49
			Pro					Pro			CAG Gln 5		TTG Leu	97
	Thr					Cys					Asp		ACA Thr 25	145
					Pro					Asp	TTC Phe		Asn	193
				Asp							CTT Leu			241
			Phe								CAG G1n			289
											CTG Leu 85			337
	Ala										GAC Asp			385
											ACC Thr			433
											TTT Phe			481
											GCC Ala			529
											AGG Arg 165			577
											ACC Thr			625

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CAC His	CCC Pro	GAG Glu	GAG G lu	CTG Leu 190	GTG Val	CTG Leu	CTC Leu	GGA Gly	CAC His 195	TCT Ser	CTG Leu	GGC Gly	ATC Ile	CCC Pro 200	TGG Trp	673
GCT Ala	CCC Pro	CTG Leu	AGC Ser 205	TCC Ser	TGC Cys	CCC Pro	AGC Ser	CAG Gln 210	GCC Ala	CTG Leu	CAG Gln	CTG Leu	GCA Ala 215	GGC Gly	TGC Cys	721
TTG Leu	AGC Ser	CAA G1n 220	CTC Leu	CAT His	AGC Ser	GGC Gly	CTT Leu 225	TTC Phe	CTC Leu	TAC Tyr	CAG Gln	GGG Gly 230	CTC Leu	CTG Leu	CAG G In	769
Ala													CAC His			817
CAG Gln 250	CTG Leu	GAC Asp	GTC Val	GCC Ala	GAC Asp 255	TTT Phe	GCC Ala	ACC Thr	ACC Thr	ATC Ile 260	TGG Trp	CAG Gln	CAG Gln	ATG Met	GAA G lu 265	865
GAA Glu	CTG Leu	GGA Gly	ATG Met	GCC Ala 270	CCT Pro	GCC Ala	CTG Leu	CAG Gln	CCC Pro 275	ACC Thr	CAG Gln	GGT Gly	GCC Ala	ATG Met 280	CCG Pro	913
GCC Ala	TTC Phe	GCC Ala	TCT Ser 285	GCT Ala	TTC Phe	CAG Gln	CGC Arg	CGG Arg 290	GCA Ala	GGA Gly	GGG Gly	GTC Val	CTG Leu 295	GTT Val	GCT Ala	961
AGC Ser	CAT His	CTG Leu 300	CAG Gln	AGC Ser	TTC Phe	CTG Leu	GAG Glu 305	GTG Val	TCG Ser	TAC Tyr	CGC Arg	GTT Val 310	CTA Leu	CGC Arg	CAC His	1009
	GCG Ala 315			TGAT	[AAG6	AT (CGA	ATTC								1039

(2) INFORMATION FOR SEQ ID NO:14:

- (i) SEQUENCE CHARACTERISTICS:

 (A) LENGTH: 1051 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: both
- (ii) MOLECULE TYPE: DNA (genomic)
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (ix) FEATURE:

(A) NAME/KEY: CDS
(B) LOCATION: 14..1033

(ix) FEATURE:
 (A) NAME/KEY: mat_peptide
 (B) LOCATION: 71..1033

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

	(X 1) 251	ŲUEINI	ים של	L JUK.	17111	JI1	stų.	ווי פו	J.17	•					
AAT	TGCC	GCC /		ATG / Met :	AGC (Ser /	CGC (Arg l	_eu {	ccc (Pro V	STC (Val l	CTG (Leu l	CTC (Leu l	_eu l	CTC (Leu (-10	CAA (Gln l	CTC Leu	49
CTG Leu	GTC Val	CGC Arg -5	CCC Pro	GGA Gly	CTC Leu	C/\A G I n	GCT Ala 1	CCC Pro	ATG Met	ACC Thr	CAG Gln 5	ACA Thr	ACT Thr	AGT Ser	TTG Leu	97
#AG Lys 10	ACA Thr	AGC Ser	TGG Trp	GTT Val	AAC Asn 15	TGC Cys	TCT Ser	AAC Asn	ATG Met	ATC Ile 20	GAT Asp	GAA Glu	ATT Ile	ATA Ile	ACA Thr 25	145
CAC His	TTA Leu	AAC Asn	GAG Glu	CCA Pro 30	CCT Pro	TTG Leu	CCT Pro	TTG Leu	CTG Leu 35	GAC Asp	TTC Phe	AAC Asn	AAC Asn	CTC Leu 40	AAT Asn	193
GGG Gly	GAA G lu	GAC Asp	CAA Gln 45	GAC Asp	ATT Ile	CTG Leu	ATG Met	GAA Glu 50	AAT Asn	AAC Asn	CTT Leu	CGA Arg	AGG Arg 55	CCA Pro	AAC Asn	241
CTG Leu	GAG Glu	GCA Ala 60	TTC Phe	AAC Asn	AGG Arg	GCT Ala	GTC Val 65	AAG Lys	AGT Ser	TTA Leu	CAG Gln	AAT Asn 70	GCA Ala	TCA Ser	GCA Ala	289
ATT Ile	GAG Glu 75	AGC Ser	ATT Ile	CTT Leu	AAA Lys	AAT Asn 80	Leu	CTG Leu	CCA Pro	TGT Cys	CTG Leu 85	CCC Pro	CTG Leu	GCC Ala	ACG Thr	337
GCC Ala 90	GCA Ala	CCC Pro	ACG Thr	CGA Arg	CAT His 95	CCA Pro	ATC Ile	CAT His	ATC Ile	AAG Lys 100	GAC Asp	GGT Gly	GAC Asp	TGG Trp	AAT Asn 105	385
GAA 61u	TTC Phe	CGG Arg	AGG Arg	AAA Lys 110	CTG Leu	ACG Thr	TTC Phe	TAT Tyr	CTG Leu 115	AAA Lys	ACC Thr	CTT Leu	GAG Glu	AAT Asn 120	GCG Ala	433
CAG Gln	GCT Ala	CAA Gln	CAG Gln 125	ACG Thr	ACT Thr	TTG Leu	TCG Ser	CTA Leu 130	GCG Ala	TCC Ser	GGA Gly	GGC Gly	GGT Gly 135	GGC Gly	TCG Ser	481
GGC Gly	GGT Gly	GGC Gly 140	GGC Gly	TCG Ser	GGT Gly	GGC Gly	GGC Gly 145	GGC Gly	TCT Ser	GCG Ala	CTA Leu	GCG Ala 150	ATC Ile	TTT Phe	CTA Leu	529

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											CGA Arg 165					577
TAC Tyr 170	CTC Leu	TTG Leu	GAG Glu	GCC Ala	AAG Lys 175	GAG Glu	GCC Ala	GAG Glu	AAT Asn	ATC Ile 180	ACG Thr	ACG Thr	GGC Gly	TGT Cys	GCT Ala 185	625
											CCA Pro					673
											CAG Gln					721
											GTC Val					769
											CCC Pro 245					817
											ACC Thr					865
											CCA Pro					913
											TTC Phe					961
											AAG Lys					1009
	GCA Ala 315							TGAT	raag(GAT (CGA	ATTC				1051

(2) INFORMATION FOR SEQ ID NO:15:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1072 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: both
- (ii) MOLECULE TYPE: DNA (genomic)

433

	(iii) HY	POTH	ETIC	AL: I	40										
	(iv) AN	TI-SI	ENSE	: NO											
	(ix) FE/ (/ (1	A) N	AME/I	KEY: ION:	CDS 8	1054									
	(ix) FE/ (/ (!	4) N/	AME/I	KEY: ION:	mat_ 89.	_pep1 .1054	tide 4								
	(xi) SE(QUEN	CE DE	ESCR	IPTIO)N: S	SEQ :	ID NO	0:15	•					
TCGA	AGCC	ATG Met -27	GGG Gly	GTG Val -25	CAC His	GAA G lu	TGT Cys	CCT Pro	GCC Ala -20	TGG Trp	CTG Leu	TGG Trp	CTT Leu	CTC Leu -15	CTG Leu	49
TCC Ser	CTG Leu	CTG Leu	TCG Ser -10	CTC Leu	CCT Pro	CTG Leu	GGC Gly	CTC Leu -5	CCA Pro	GTC Val	CTG Leu	GGC Gly	GCC Ala l	CCA Pro	CCA Pro	97
CGC Arg	CTC Leu 5	ATC Ile	TGT Cys	GAC Asp	AGC Ser	CGA Arg 10	GTC Val	CTG Leu	GAG Glu	AGG Arg	TAC Tyr 15	CTC Leu	TTG Leu	GAG Glu	GCC Ala	145
AAG Lys 20	GAG G1u	GCC Ala	GAG Glu	AAT Asn	ATC Ile 25	ACG Thr	ACG Thr	GGC Gly	TGT Cys	GCT Ala 30	GAA G 1u	CAC His	TGC Cys	AGC Ser	TTG Leu 35	193
AAT Asn	GAG Glu	AAT Asn	ATC Ile	ACT Thr 40	GTC Val	CCA Pro	GAC Asp	ACC Thr	AAA Lys 45	GTT Val	AAT Asn	TTC Phe	TAC Tyr	GCG Ala 50	TGG Trp	241
AAG Lys	AGG A rg	ATG Met	GAG Glu 55	GTC Val	GGC Gly	CAG Gln	CAG Gln	GCC Ala 60	GTA Val	GAA Glu	GTC Val	TGG Trp	CAG Gln 65	GGC Gly	CTG Leu	289
GCC Ala	CTG Leu	CTG Leu 70	TCG Ser	GAA G lu	GCT Ala	GTC Val	CTG Leu 75	CGG Arg	GGC Gly	CAG Gln	GCC Ala	CTG Leu 80	TTG Leu	GTC Val	AAC Asn	337
Ser	Ser	CAG Gln	Pro	Trp	Glu	Pro	Leu	Gln	CTG Leu	His	GTG Val 95	GAT Asp	AAA Lys	GCC Ala	GTC Val	385

AGT GGC CTT CGC AGC CTC ACC ACT CTG CTT CGG GCT CTG GGA GCT CAG Ser Gly Leu Arg Ser Leu Thr Thr Leu Leu Arg Ala Leu Gly Ala Gln 100 105 110

- 52 -

l	AG .ys	GAA G Tu	GCC Ala	ATC Ile	TCC Ser 120	CCT Pro	CCA Pro	GAT Asp	GCG Ala	GCC Ala 125	TCA Ser	GCT Ala	GCT Ala	CCA Pro	CTC Leu 130	CGA Arg	481
1	ACA Thr	ATC Ile	ACT Thr	GCT Ala 135	GAC Asp	ACT Thr	TTC Phe	CGC Arg	AAA Lys 140	CTC Leu	TTC Phe	CGA Arg	GTC Val	TAC Tyr 145	TCC Ser	AAT Asn	529
1	TTC he	CTC Leu	CGG Arg 150	GGA Gly	AAG Lys	CTG Leu	AAG Lys	CTG Leu 155	TAC Tyr	ACA Thr	GGG Gly	GAG G lu	GCA Ala 160	TGC Cys	AGG Arg	ACA Thr	577
(GGG 31y	GAC Asp 165	AGA Arg	GCG Ala	GCC Ala	GCC Ala	TCC Ser 170	GGA Gly	GGC Gly	GGT Gly	GGC Gly	TCG Ser 175	GGC Gly	GGT Gly	GGC Gly	GGC Gly	625
9	CG Ser 180	GGT Gly	GGC Gly	GGC Gly	GGC Gly	TCT Ser 185	GCG Ala	GCC Ala	GCC Ala	ATG Met	GCT Ala 190	CCC Pro	ATG Met	ACC Thr	CAG Gln	ACA Thr 195	673
į	ACT Tar	AGT Ser	TTG Leu	AAG Lys	ACA Thr 200	AGC Ser	TGG Trp	GTT Val	AAC Asn	TGC Cys 205	TCT Ser	AAC Asn	ATG Met	ATC Ile	GAT Asp 210	GAA Glu	721
	ATT I le	ATA I le	ACA Thr	CAC His 215	TTA Leu	AAC Asn	GAG G1u	CCA Pro	CCT Pro 220	TTG Leu	CCT Pro	TTG Leu	CTG Leu	GAC Asp 225	TTC Phe	AAC Asn	769
1	AAC Asn	CTC Leu	AAT Asn 230	GGG Gly	GAA G lu	GAC Asp	CAA Gln	GAC Asp 235	ATT Ile	CTG Leu	ATG Met	GAA G lu	AAT Asn 240	AAC Asn	CTT Leu	CGA Arg	817
4	AGG Arg	CCA Pro 245	AAC Asn	CTG Leu	GAG G lu	GCA Ala	TTC Phe 250	AAC Asn	AGG Arg	GCT Ala	GTC Val	AAG Lys 255	AGT Ser	TTA Leu	CAG Gln	AAT Asn	865
	GCA Ala 260	TCA Ser	GCA Ala	ATT Ile	GAG G lu	AGC Ser 265	Ile	CTT Leu	AAA ·Lys	AAT Asn	CTC Leu 270	CTG Leu	CCA Pro	TGT	CTG Leu	CCC Pro 275	913
	CTG Leu	GCC Ala	ACG Thr	GCC Ala	GCA Ala 280	Pro	ACG Thr	CGA Arg	CAT His	CCA Pro 285	Ile	CAT His	ATC Ile	AAG Lys	GAC Asp 290	Gly	961
	GAC Asp	TGG Trp	AAT Asn	GAA G lu 295	Phe	CGG Arg	AGG Arg	AAA Lys	CTG Leu 300	Thr	TTC Phe	TAT Tyr	CTG Leu	Lys 305	Thr	CTT Leu	1009
	GAG Glu	AAT Asn	GCG Ala 310	Gln	GCT Ala	CAA G1n	CAG Gln	ACG Thr 315	Thr	TTG Leu	TCG Ser	CTA Leu	GCG Ala 320	. Ile	TTT Phe		1054
	TAG	TAAG	GAT	CCGA	ATTC												1072

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(2) INFORMATION FOR SEQ ID NO:10:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 429 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: both 	
(ii) MOLECULE TYPE: DNA (genomic)	
(iii) HYPOTHETICAL: NO	
(iv) ANTI-SENSE: NO	
(ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 10411	
<pre>(ix) FEATURE: (A) NAME/KEY: mat_peptide (B) LOCATION: 13411</pre>	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:	
AAGCTTACC ATG GCT CCC ATG ACC CAG ACA ACT AGT TTG AAG ACA AGC Met Ala Pro Met Thr Gln Thr Thr Ser Leu Lys Thr Ser -1 1 5 10	48
TGG GTT AAC TGC TCT AAC ATG ATC GAT GAA ATT ATA ACA CAC TTA AAC Trp Val Asn Cys Ser Asn Met Ile Asp Glu Ile Ile Thr His Leu Asn 15 20 25	96
GAG CCA CCT TTG CCT TTG CTG GAC TTC AAC AAC CTC AAT GGG GAA GAC Glu Pro Pro Leu Pro Leu Leu Asp Phe Asn Asn Leu Asn Gly Glu Asp 30 35.	144
CAA GAC ATT CTG ATG GAA AAT AAC CTT CGA AGG CCA AAC CTG GAG GCA Gln Asp Ile Leu Met Glu Asn Asn Leu Arg Arg Pro Asn Leu Glu Ala 45 50 55 60	192
TTC AAC AGG GCT GTC AAG AGT TTA CAG AAT GCA TCA GCA ATT GAG AGC Phe Asn Arg Ala Val Lys Ser Leu Gln Asn Ala Ser Ala Ile Glu Ser 65 70 75	240
ATT CTT AAA AAT CTC CTG CCA TGT CTG CCC CTG GCC ACG GCC GCA CCC Ile Leu Lys Asn Leu Leu Pro Cys Leu Pro Leu Ala Thr Ala Ala Pro 80 85 90	288
ACG CGA CAT CCA ATC CAT ATC AAG GAC GGT GAC TGG AAT GAA TTC CGG Thr Arg His Pro Ile His Ile Lys Asp Gly Asp Trp Asn Glu Phe Arg 95 100 105	336

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AGG AAA CTG ACG TTC TAT CTG AAA ACC CTT GAG AAT GCG CAG GCT CAA Arg Lys Leu Thr Phe Tyr Leu Lys Thr Leu Glu Asn Ala Gln Ala Gln 110 115 120	384
CAG ACG ACT TTG TCG CTA GCG ATC TTT TAGTAAGGAT CCGAATTC Gln Thr Thr Leu Ser Leu Ala Ile Phe 125 130	429
(2) INFORMATION FOR SEQ ID NO:17:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 532 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: double(D) TOPOLOGY: both	
(ii) MOLECULE TYPE: DNA (genomic)	
(iii) HYPOTHETICAL: NO	
(iv) ANTI-SENSE: NO	
(ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 14514	
<pre>(ix) FEATURE: (A) NAME/KEY: mat_peptide (B) LOCATION: 17514</pre>	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:	
AAGCTTACCT GCC ATG GCC CCA CCA CGC CTC ATC TGT GAC AGC CGA GTC Met Ala Pro Pro Arg Leu Ile Cys Asp Ser Arg Val -1 1 5 10	49
CTG GAG AGG TAC CTC TTG GAG GCC AAG GAG GCC GAG AAT ATC ACG ACG Leu Glu Arg Tyr Leu Leu Glu Ala Lys Glu Ala Glu Asn Ile Thr Thr	97
GGC TGT GCT GAA CAC TGC AGC TTG AAT GAG AAT ATC ACT GTC CCA GAC Gly Cys Ala Glu His Cys Ser Leu Asn Glu Asn Ile Thr Val Pro Asp 30 35 40	145
ACC AAA GTT AAT TTC TAC GCG TGG AAG AGG ATG GAG GTC GGC CAG CAG Thr Lys Val Asn Phe Tyr Ala Trp Lys Arg Met Glu Val Gly Gln Gln 45 50 55	193
GCC GTA GAA GTC TGG CAG GGC CTG GCC CTG CTG TCG GAA GCT GTC CTG Ala Val Glu Val Trp Gln Gly Leu Ala Leu Leu Ser Glu Ala Val Leu 60 65 70 75	241

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CGG Arg	GGC Gly	CAG Gln	GCC Ala	CTG Leu 80	TTG Leu	GTC Val	AAC Asn	TCG Ser	AGC Ser 85	CAG Gln	CCG Pro	TGG Trp	GAG Glu	CCC Pro 90	CTG Leu	289
CAA G1n	CTG Leu	CAT His	GTG Val 95	GAT Asp	AAA Lys	GCC Ala	GTC Val	AGT Ser 100	GGC Gly	CTT Leu	CGC A rg	AGC Ser	CTC Leu 105	ACC Thr	ACT Thr	337
CTG Leu	CTT Leu	CGG Arg 110	GCT Ala	CTG Leu	GGA Gly	GCT Ala	CAG Gln 115	AAG Lys	GAA Glu	GCC Ala	ATC Ile	TCC Ser 120	CCT Pro	CCA Pro	GAT Asp	385
											GCT Ala 135					433
AAA Lys 140	CTC Leu	TTC Phe	CGA Arg	GTC Val	TAC Tyr 145	TCC Ser	AAT Asn	TTC Phe	CTC Leu	CGG Arg 150	GGA Gly	AAG Lys	CTG Leu	AAG Lys	CTG Leu 155	481
				GCA Ala 160						AG A	TGAT	AAG	A TO	CGAA	ATTC	532

(2) INFORMATION FOR SEQ ID NO:18:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 556 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: both
- (ii) MOLECULE TYPE: DNA (genomic)
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (ix) FEATURE:

 - (A) NAME/KEY: CDS (B) LOCATION: 14..538
- (ix) FEATURE:
 - (A) NAME/KEY: mat_peptide (B) LOCATION: 17..538
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

AAGCTTACCT GCC ATG ACC CCC CTG GGC CCT GCC AGC TCC CTG CCC CAG 49 Met Thr Pro Leu Gly Pro Ala Ser Ser Leu Pro Gln

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									CAG Gln 25		97
									CTG Leu		145
									CCC Pro		193
									GGC Gly		241
									CTG Leu		289
									ACA Thr 105		337
									ATG Met		385
									ATG Met		433
									GTT Val		481
									CGC Arg		529
	CAG Gln	TGA	TAAG	GAT (CCGA	ATTC					556

(2) INFORMATION FOR SEQ ID NO:19:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 39 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:	
AATTGCCGCC ACCATGAGCC GCCTGCCCGT CCTGCTCCT	39
(2) INFORMATION FOR SEQ ID NO:20:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 50 hase pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA	
(iii) HYPOTHETICAL: NO	
(iv) ANTI-SENSE: NO	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20: GCTCCAACTC CTGGTCCGCC CCGGACTCCA AGCTCCCATG ACCCAGACAA (2) INFORMATION FOR SEQ ID NO:21. (i) SEQUENCE CHARACTERISTICS:	50
(A) LENGTH: 39 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA	
(iii) HYPOTHETICAL: NO	
(iv) ANTI-SENSE: NO	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:	
CTAGTTGTCT GGGTCATGGG AGCTTGGAGT CCGGGGCGG	39

(2) INFORMATION FOR SEQ ID NO:22:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 50 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	·
(ii) MOLECULE TYPE: DNA	
(iii) HYPOTHETICAL: NO	
(iv) ANTI-SENSE: NO	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:	
ACCAGGAGTT GGAGCAGGAG CAGGACGGCC AGGCGGCTCA TGGTGGCGGC	50
(2) INFORMATION FOR SEQ ID NO:23:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 17 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(ii) MOLECULE TYPE: DNA	
(iii) HYPOTHETICAL: NO	
(iv) ANTI-SENSE: NO	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:	
CTAGCGATCT TTCTAGA	17
(2) INFORMATION FOR SEQ ID NO:24:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 17 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(ii) MOLECULE TYPE: DNA	
(iii) HYPOTHETICAL: NO	

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(iv) ANTI-SENSE: NO	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:	
CATGTCTAGA AAGATCG	17
(2) INFORMATION FOR SEQ ID NO:25:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 57 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(ii) MOLECULE TYPE: DNA	
(iii) HYPOTHETICAL: NO	
(iv) ANTI-SENSE: NO	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:	
CTAGCGTCCG GAGGCGGTGG CTCGGGCGGT GGCGGCTCGG GTGGCGGCGG CTCTGCG	57
(2) INFORMATION FOR SEQ ID NO:26:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 57 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA	
(iii) HYPOTHETICAL: NO	
(iv) ANTI-SENSE: NO	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:	
CTAGCGCAGA GCCGCCGCCA CCGCAGCCGC CACCGCCCGA GCCACCGCCT CCGGACG	57
(2) INFORMATION FOR SEQ ID NO:27:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 15 amino acids	

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- (B) TYPE: amino acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:

- (2) INFORMATION FOR SEQ ID NO:28:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 21 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA
 - (iii) HYPOTHETICAL: NO
 - (iv) ANTI-SENSE: NO
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:

TTGTCGCTAG CGTCCGGAGG C

21

- (2) INFORMATION FOR SEQ ID NO:29:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 15 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA
 - (iii) HYPOTHETICAL: NO
 - (iv) ANTI-SENSE: NO

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:	
AAGCGG CCGCA	15
INFORMATION FOR SEQ ID NO:30:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 15 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA	
iii) HYPOTHETICAL: NO	
(iv) ANTI-SENSE: NO	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:	
CCGGCG TTCGA	15
INFORMATION FOR SEQ ID NO:31:	
 (i) SEQUENCE CHARACTERISTICS: (A)-LENGTH: 28 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(ii) MOLECULE TYPE: DNA	
iii) HYPOTHETICAL: NO	
(iv) ANTI-SENSE: NO	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:31:	
GCCATG GGGGTGCACG AATGTCCT	28
INFORMATION FOR SEQ ID NO:32:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 32 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	

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(ii) MOLECULE TYPE: DNA	
(iii) HYPOTHETICAL: NO	
(iv) ANTI-SENSE: NO	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:	
GCCTGGCTGT GGCTTCTCCT GTCCCTGCTG TC	32
(2) INFORMATION FOR SEQ ID NO:33:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 32 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(ii) MOLECULE TYPE: DNA	
(iii) HYPOTHETICAL: NO	
(iv) ANTI-SENSE: NO	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:	
GCTCCCTCTG GGCCTCCCAG TCCTGGGCTG CA	32
(2) INFORMATION FOR SEQ ID NO:34:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 26 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA	
(iii) HYPOTHETICAL: NO	
(iv) ANTI-SENSE: NO	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:34:	
GCCCAGGACT GGGAGGCCCA GAGGGA	26

(2) INFORMATION FOR SEQ ID NO:35:

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(A) LENG (B) TYPI (C) STR/	CHARACTERISTICS: GTH: 42 base pairs E: nucleic acid ANDEDNESS: single OLOGY: linear	
(ii) MOLECULE	TYPE: DNA	
(iii) HYPOTHET	ICAL: NO	
(iv) ANTI-SENS	SE: NO	
COLON CEONEMOS	DECCRIPTION, CEO ID MO.25.	
•	DESCRIPTION: SEQ ID NO:35:	42
	GAGA AGCCACAGCC AGGCAGGACA TT	42
(2) INFORMATION FO	OR SEQ ID NO:36:	
(A) LENG (B) TYPE (C) STRA	CHARACTERISTICS: GTH: 16 base pairs E: nucleic acid ANDEDNESS: single DLOGY: linear	
(ii) MOLECULE	TYPE: DNA	
(iii) HYPOTHETI	ICAL: NO	
(iv) ANTI-SENS	SE: NO	
(xi) SEQUENCE	DESCRIPTION: SEQ ID NO:36:	
CGTGCACCCC CATGGC		16
(2) INFORMATION FO	OR SEQ ID NO:37:	
(A) LENG (B) TYPE (C) STRA	CHARACTERISTICS: GTH: 21 base pairs E: nucleic acid ANDEDNESS: single DLOGY: linear	
(ii) MOLECULE	TYPE: DNA	
(iii) HYPOTHETI	ICAL: NO	
(iv) ANTI-SENS	SE: NO	

(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:37:	
GCCCCACCA	AC GCCTCATCTG T	21
(2) INFO	RMATION FOR SEQ ID NO:38:	
(i)	SEQUENCE CHARACTERISTICS: (A) LENGTH: 21 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii)	MOLECULE TYPE: DNA	
(iii)	HYPOTHETICAL: NO	
(iv)	ANTI-SENSE: NO	
(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:38:	
GAATTCGGA	AT CCTTATCATC T	21
(0) 14505		
(2) INFOR	RMATION FOR SEQ ID NO:39:	
(i)	SEQUENCE CHARACTERISTICS: (A) LENGTH: 30 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii)	MOLECULE TYPE: DNA	
(iii)	HYPOTHETICAL: NO	
(iv)	ANTI-SENSE: NO	
(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:39:	
CTAGTCTCT	TA GAATGGGGGT CCACGAATGT	30
(2) INFOR	RMATION FOR SEQ ID NO:40:	
(i)	SEQUENCE CHARACTERISTICS: (A) LENGTH: 30 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	

(ii) MOLECULE TYPE: DNA

- 65	-
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(iii) HYPOTHETICAL: NO	
(iv) ANTI-SENSE: NO	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:40:	
AGCCATGGCG GCCGCTCTGT CCCCTGTCCT	30
(2) INFORMATION FOR SEQ ID NO:41:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 30 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA	•
(iii) HYPOTHETICAL: NO	
(iv) ANTI-SENSE: NO	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:41:	
GACAGAGCGG CCGCCATGGC TCCCATGACC	30
(2) INFORMATION FOR SEQ ID NO:42:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 30 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA	
(iii) HYPOTHETICAL: NO	
(iv) ANTI-SENSE: NO	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:42:	30

(2)	INFORMATION	FOR	SEQ	ID	NO:4	13:
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- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 57 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:43:

GGCCGCTTCC GGAGGCGGTG GCTCGGGCGG TGGCGGCTCG GGTGGCGGCG GCTCTGC

57

- (2) INFORMATION FOR SEQ ID NO:44:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 57 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA
 - (iii) HYPOTHETICAL: NO
 - (iv) ANTI-SENSE: NO
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:44:

GGCCGCAGAG CCGCCGCCAC CCGAGCCGCC ACCGCCCGAG CCACCGCCTC CGGCAGC

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What is claimed is:

- 1. A recombinant hematopoietic molecule comprising at least a portion of a first hematopoietic molecule having early myeloid differentiation activity and at least a portion of a second hematopoietic molecule having late myeloid differentiation activity, said recombinant hematopoietic molecule having early myeloid differentiation activity associated with said first hematopoietic molecule and late myeloid differentiation activity associated with said second hematopoietic molecule.
- 2. A recombinant hematopoietic molecule of claim 1 wherein the first hematopoietic molecule is selected from the group consisting of IL-3 and GM-CSF.
- 3. A recombinant hematopoietic molecule of claim 1 wherein the second hematopoietic molecule is selected from the group consisting of Epo, G-CSF, IL-5 and M-CSF.
- 4. A recombinant hematopoietic molecule of claim 1 wherein the portion of the first hematopoietic molecule is linked to the portion of the second hematopoietic molecule by an amino acid linker sequence of at least two amino acid residues.
 - 5. A recombinant hematopoietic molecule of claim 1 comprising SEQ ID NO: 1.
 - 6. A recombinant hematopoietic molecule of claim 1 comprising an amino acid sequence contained within SEQ ID NO: 2.
 - 7. A recombinant hematopoietic molecule of claim 1 comprising SEQ ID NO: 3.
- 8. A recombinant hematopoietic molecule of claim 1 comprising an amino acid sequence contained within SEQ ID NO: 4.

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- 9. A recombinant hematopoietic molecule of claim 1 comprising SEQ ID NO: 5.
- IO. A recombinant hematopoietic molecule of claim 1 wherein the first hematopoietic molecule is IL-3 and the second hematopoietic molecule is Epo.
 - 11. A recombinant hematopoietic molecule of claim 10 wherein the first hematopoietic molecule comprises the amino portion and the second hematopoietic molecule comprises the carboxy portion of the recombinant hematopoietic molecule.

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12. A recombinant hematopoietic molecule of claim 11 which comprises SEQ ID NO: 6.

13. A recombinant hematopoietic molecule of claim 11 which comprises SEQ ID NO: 7.

- 14. A recombinant hematopoietic molecule of claim 10 wherein the first hematopoietic molecule comprises the carboxy portion and the second hematopoietic molecule comprises the amino portion of the recombinant hematopoietic molecule.
- 15. A recombinant hematopoietic molecule of claim 14 which comprises SEQ ID NO: 8.
 - 16. A recombinant hematopoietic molecule of claim 14 which comprises SEQ ID NO: 9.
- 17. A recombinant hematopoietic molecule of claim 1 wherein the first hematopoietic molecule is IL-3 and the second hematopoietic molecule is G-CSF

18. A recombinant hematopoietic molecule of claim 17 wherein the first hematopoietic molecule comprises the amino portion and the second hematopoietic molecule comprises the carboxy portion of the recombinant hematopoietic molecule.

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- 19. A recombinant hematopoietic molecule of claim 18 which comprises SEQ ID NO: 10.
- 20. A nucleic acid molecule which encodes the recombinant hematopoietic molecule of claim 1.
 - 21. An expression vector which comprises the nucleic acid molecule of claim 20.
- 15 22. A host cell transformed with the expression vector of claim
 19.
 - 23. A host cell of claim 22 which comprises a mammalian cell.

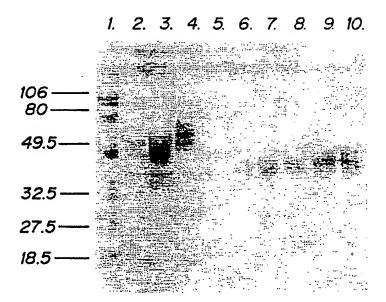
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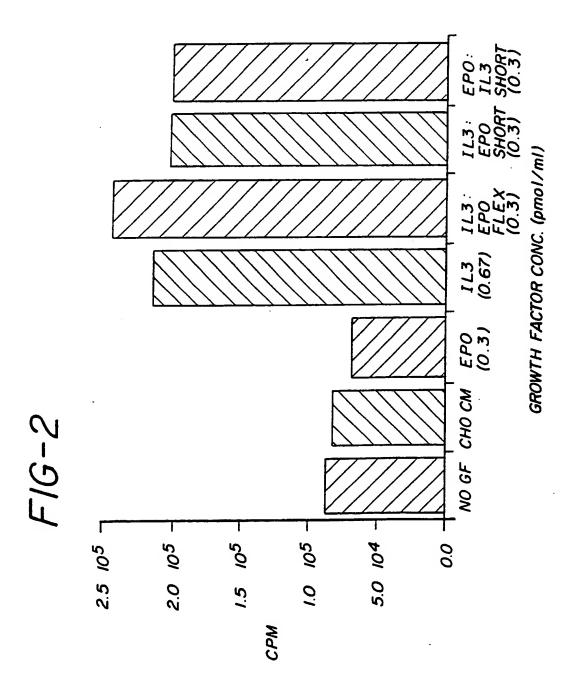
25

- 24. A method for producing a recombinant hematopoietic molecule comprising at least a portion of a first hematopoietic molecule having early myeloid differentiation activity and at least a portion of a second hematopoietic molecule having late myeloid differentiation activity, which comprises culturing a host cell of claim 22 under suitable conditions so as to allow the expression of such recombinant hematopoietic molecule, and recovering such recombinant hematopoietic molecule.
- 25. A pharmaceutical composition which comprises a recombinant hematopoietic molecule of claim 1 and a pharmaceutically acceptable carrier.
- 26. A method for promoting hematopoiesis in a patient which comprises administering to such patient a pharmaceutical composition of claim 25.

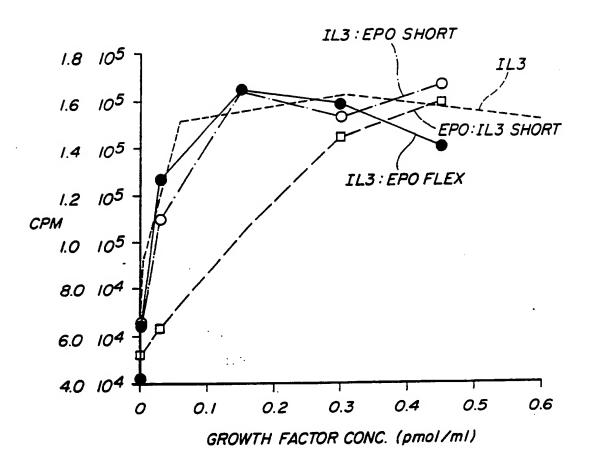
1/11

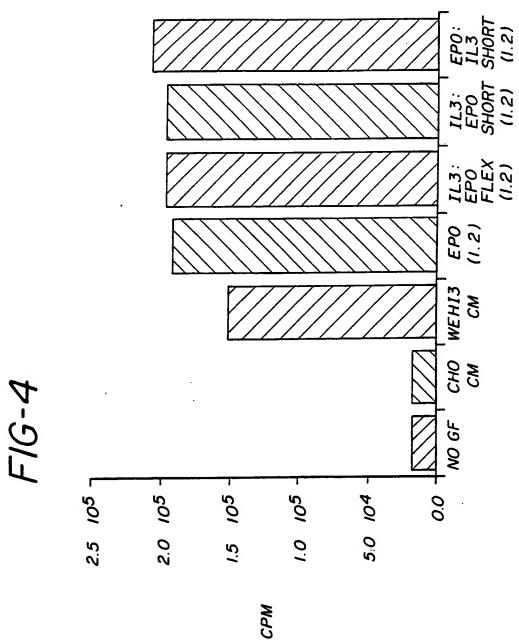
FIG-1





F/G-3





GROWTH FACTOR CONC. (pmol/ml)

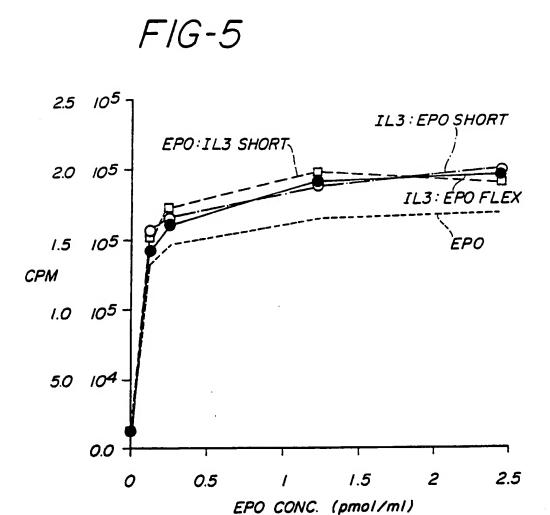
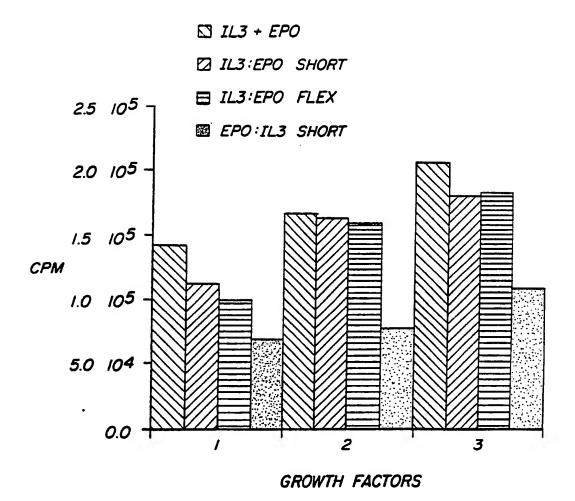
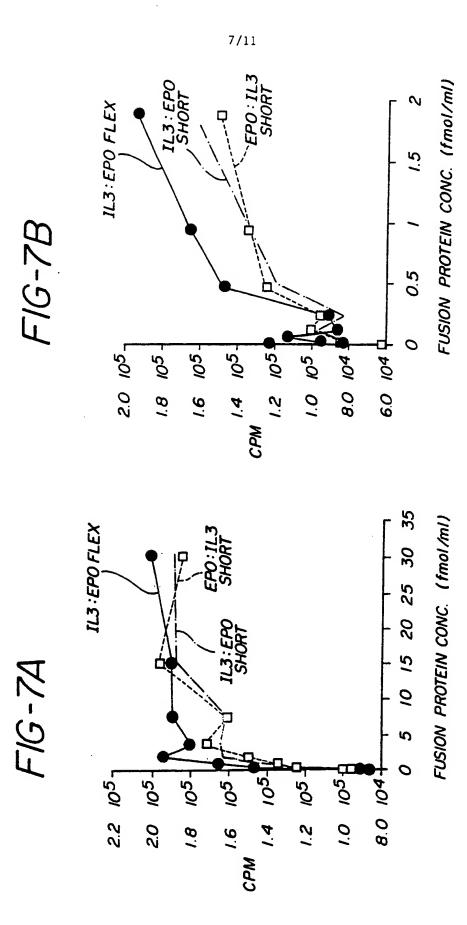
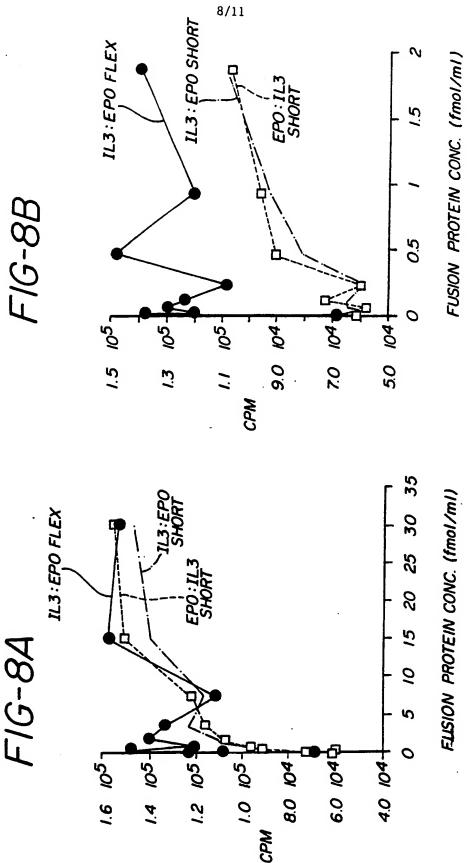
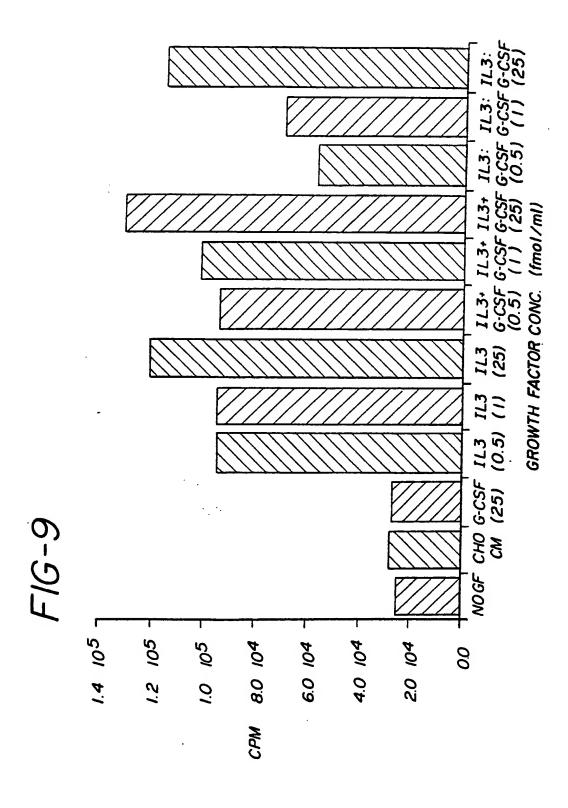


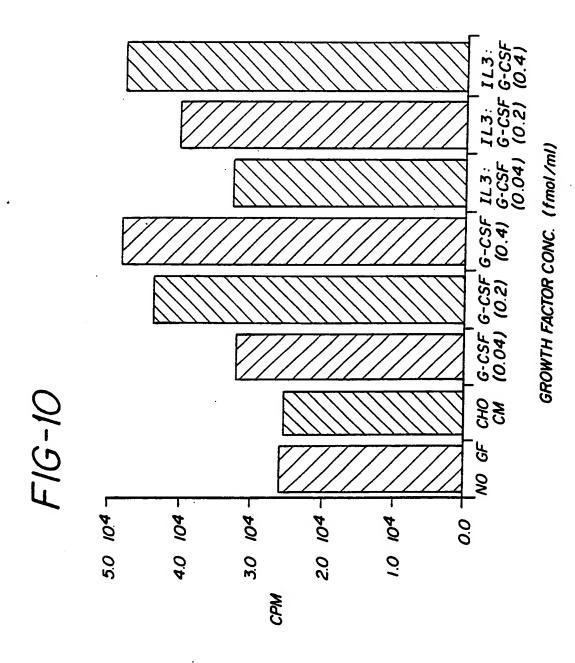
FIG-6

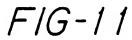


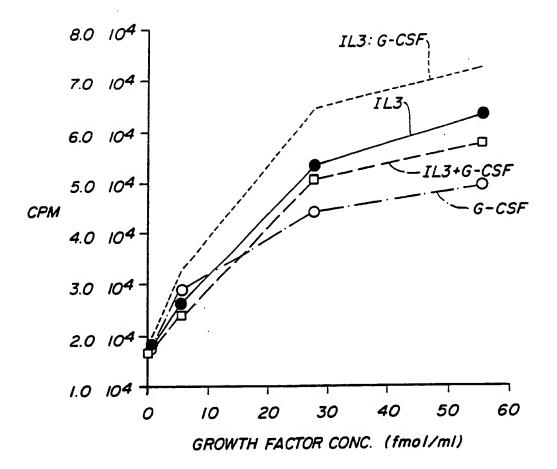












INTERNATIONAL SEARCH REPORT

International Application No. PCT/US91/07053

International Application No. PCT/US91/07053						
I. CLASSI	FICATIO	N OF SUBJECT MATTER (if several class	sification symbols apply, indicate all) 4			
U.S .CL.	: 530/3	onal Patent Classification (IPC) or to both N 20; C12P21/02; C12N 15/24, 15/ 51,395;435/69.5,69.51,69.6,69.	27, 15/70, 5/00,5/10; A61K 37	/02; CO7H 15/12 ; 536/27		
II. FIELDS	SEARCH	ED				
		Minimum Docum	entation Searched ?			
Classification	n System		Classification Symbols			
U.S.	U.S. 530/350,351,395; 435/69.1,69.5,69.51, 69.52, 69.6,69.7,172.3 240.1,252.3, 320.1; 424/85.1,85.2; 536/27					
			r than Minimum Documentation ts are included in the Fields Searched ⁸			
Computer	r Data-	-base search				
III. DOCUM	MENTS C	INSIDERED TO BE RELEVANT				
Category *	Citatio	in of Document, 11 with indication, where as	propriate, of the relevant passages 12	Relevant to Claim No. 13		
$E, \frac{X}{Y}$	US. 17 D	A. 5.073.627 (Curtis ecember 1991, see cl	et al.) aims.	1-2.20-26,4 1-26		
ï	US. 19 J clai	A. 4.935.233 (Bell e une 1990, see col. 3 ms.	t al.) -4 and	1-26		
ï	US 19 Ju	A. 4.935,352 (Koichi une 1990, see all.	et al.)	1-26		
P.Y	WO. 3	A. 91/01004 (Svrluga anuary 1991. see all.	et al.)	1-26		
Α	US. 3 23 Ju	. 4.675.382 (Murphy) one 1987. see all.		1-26		
Y	WO. A 11 Fe	, 88/00971 (Ramshaw bruary 1988, see all	et al.)	1-26		
			-	•		
* Special categories of cited documents: 10 "A" document defining the general state of the art which is not considered to be of particular relevance in the application out considered to be of particular relevance in the international filling date "E" earner document but published on or after the international filling date "L" document which may throw doubts on priority claim(s) or which is cried to establish the publication date of another citation or other special reason (as specified) "O" document referring to an oral disclosure, use, exhibition or other means "P" document published prior to the international filling date but later than the priority date claimed IV. CERTIFICATION Date of the Actual Completion of the international Search O? FEB 1992						
International Searching Authority ISA/US Signature of Authorized Search Garnette D. Draper ebw						